

Mitochondrial DNA 4977 BP deletion mutations in lung carcinoma

Ji Gang Dai, Ying Bin Xiao, Jia Xin Min, Guo Qiang Zhang, Ke Yao, Ren Jie Zhou

Department of Thoracic Cardiovascular Surgery of Xinqiao Hospital, The Third Military Medical University, Chongqing - 400037, China.

Correspondence to: Xiao YB, E-mail: daijigangdaijigang@yahoo.co.nz

Abstract

BACKGROUND: The most common and also the most often assayed mtDNA deletion mutation, \square mtDNA⁴⁹⁷⁷ has been demonstrated in various types of human cancer. However, knowledge about \square mtDNA⁴⁹⁷⁷ in lung carcinoma is poor. **AIM:** To study the 4977 bp deletions of mitochondrial DNA (\square mtDNA⁴⁹⁷⁷) in lung cancer, adjacent histologically normal and normal lung tissue and its potential roles in the development of cancer. **MATERIALS AND METHODS:** Thirty-seven matched lung cancer/adjacent histologically normal and 20 histologically normal lung tissue samples in subjects without lung cancer were analyzed by PCR technique. **RESULTS:** \square mtDNA⁴⁹⁷⁷ deletions were detected in 54.1% (20/37) of lung cancers, 59.5% (22/37) of adjacent normal and 30.0% (6/20) of normal lung tissue samples. No significant difference was found in the frequency of \square mtDNA⁴⁹⁷⁷ deletions between the tumor and adjacent normal lung tissues (P value = 0.815). Moreover, no significant difference was found in the frequency of \square mtDNA⁴⁹⁷⁷ deletions between the tumor and histologically normal lung tissues in subjects without lung cancer (P value = 0.101). However, the correlation between \square mtDNA⁴⁹⁷⁷ deletion and age and smoking factors was present in our data. **STATISTICAL ANALYSIS:** Fisher's exact test was used to assess the difference in different groups by the Scientific Package for Social Sciences (SPSS), version 10.0, Statistical analysis software. **CONCLUSIONS:** Mitochondrial DNA 4977 bp deletion, which is not specific to lung cancer, may reflect the environmental and aging process influences operative during tumor progression.

Key Words: Deletion, lung carcinoma, mitochondrial DNA, mutation

Mitochondria are the intracellular organelles responsible for adenosine triphosphate (ATP) synthesis through the coupling of oxidative phosphorylation (OXPHOS) to mitochondrial respiration in human and animal cells. They contain their own genome in the form of mitochondrial DNA (mtDNA), which is the only extrachromosomal DNA in human cells. Mitochondria are involved in apoptosis and probably also tumorigenesis,^[1,2] which has led researchers to examine the potential roles of mtDNA alterations in the development and maintenance of cancers. Mammalian mitochondrial DNA (mtDNA) is a circular double-stranded DNA of 16.5 Kb in size. In contrast to the nuclear DNA, mtDNA is a naked compact DNA molecule without introns and is replicated at a much higher rate without an efficient DNA repair mechanism.

Therefore, mtDNA is more vulnerable to attacks by reactive oxygen species and free radicals. Each nucleated human cell contains a few thousand copies of mtDNA, the somatic mutation rate of which is presumed to be 10 to 20 times higher than that of nuclear DNA.^[3-5]

Human mitochondrial DNA (mtDNA) is becoming the study hotspot for its alteration in correlation with its tumorigenesis.^[6] mtDNA mutations were reported in different types of cancer and cancer cell lines. Reported sequence changes include point mutations (mostly transitions), multiple deletions and microsatellite instability in coding and noncoding regions.^[7-11] Mitochondrial DNA 4977 bp deletion (Δ mtDNA⁴⁹⁷⁷) is the most common change in mtDNA and has been detected in several types of human tumors including

gastric cancer, esophageal carcinoma, hepatocellular carcinoma, thyroid tumors, etc.^[12-14] However, little is known about this deletion of mtDNA in lung cancer. In this study, Δ mtDNA⁴⁹⁷⁷ deletion mutation was detected in lung cancer, adjacent histologically normal and histologically normal lung tissue samples in subjects without lung cancer and the correlation between Δ mtDNA⁴⁹⁷⁷ and age and smoking factors was analyzed.

Materials and Methods

Subjects

The following specimens were collected at our hospital from January 2003 to December 2003, including 37 matched lung cancer and adjacent histologically normal tissue samples and 20 normal lung tissues from patients without lung cancer. (STR typing was not done.) Patients with infectious diseases such as pneumonia, fungal infection and pulmonary tuberculosis; those with centrally located tumors with obstructing pneumonitis; and those who had received chemotherapy or radiotherapy for primary or metastatic lung malignancies were excluded from the study. None of the patients recruited into the study had a known history of industrial or occupational exposure to asbestos or organic solvent. All the tissues were kept in liquid nitrogen immediately after surgical resection according to a protocol, which were in accordance with the ethical standards of the responsible committee for conducting human research at the hospital and with the Helsinki Declaration of 1975, as revised in 2000. Smoking habit was classified into three categories: (1) current smokers, consisting of subjects who had smoked more than one cigarette or part thereof daily for more than 1 yr; (2) ex-smokers, consisting of subjects who had been previously reported as smokers and had quit smoking for more than 1 yr; and (3) nonsmokers, consisting of subjects who had never smoked. [Table 1] is a further description of the patient materials.

Total cellular DNA containing the mtDNA and nucleus DNA (nDNA) was prepared using the routine method: (1) Small amounts of tissue (100 mg) were snipped off, suspended in cell lysis buffer and incubated for 1 h at 37°C. (2) The cell suspensions were mixed by vortexing and incubated for 3 h at 50°C after the addition of enzyme K (20 mg/ml). (3) They were mixed by vortexing for 10 min and centrifuged at 5000 r/min for 15 min at room temperature after the addition of equal volume of Tris-saturated phenol. (4) The aqueous fraction was extracted twice with an equal volume of phenol: chloroform: isopentanol (25:24:1). (5) The aqueous fraction was preserved at - 20°C for 1 h after the addition of 2 volume of ethanol and 0.1

Table 1: General information of the patients

	Cancer (n=37)	Adjacent normal (n=37)	Normal (n=20)
Age (Median age)	30~75 (55)	30~75 (55)	25-70 (50)
Sex			
Female	9	9	13
Males	28	28	7
Histological types			
Squamous cell cancer	23		
Adenocarcinoma	14		
Smoking habit			
Smokers	15	15	9
Ex-smokers	9	9	4
Nonsmokers	13	13	7

volume of 3 mol/L sodium acetate. (6) Total DNA was collected as a pellet by centrifugation at 12 000 r/min for 10 min at 4°C. (7) DNA was washed once in 75% ethanol, collected by centrifugation at 12 000 r/min for 5 min and dried at room temperature. (8) DNA was resuspended in TE (10 mmol/L Tris-HCl, PH 7.5, 1 mmol/L EDTA) and stored at - 20°C.

Detection of mitochondrial common deletion

The Δ mtDNA⁴⁹⁷⁷ deletion was detected as described by Rogounovitch *et al*^[15] with two sets of primers (P1/P2 and P3/P4), one giving a product only in wild-type DNA, the other only in DNA with the Δ mtDNA⁴⁹⁷⁷ deletion. Primer sequences: P1, 5'-CTG AGC CTT TTA CCA CTC CAG-3' (nt 9500~nt 9520); P2, 5'-GGT GAT TGA TAC TCC TGA TGC GN3' (nt 9641~ nt 9619); P3, 5'-CCC ACT GTA AAG CTA ACT TAG CAT TAA CCN3' (nt 8293 ~ nt 8321); P4, 5'-GGT TTC GAT GAT GTG GTC TTT GN3' (nt 13530 ~ nt 13509).

PCR reactions were carried out using TaKaRa PCR kit (TaKaRa, Japan) in a 50 μ l reaction volume with 200 ng DNA template, 2 U TaKaRa Taq DNA polymerase (TaKaRa, Japan), 2.5 mmol/L MgCl₂, 250 μ mol/L each dNTP and 0.5 μ mol/L of each primer. After denaturation at 94°C for 30 s, the reaction mixture was cycled 30 times at 94°C for 30 s, 59°C for 30 s and 72°C for 1 min, finally extended at 72°C for 10 min.

PCR products were analyzed by 1% agarose gel electrophoresis at 60V (the buffer fluid was 1 \times TAE buffer). The electrophoresis gels were observed under ultraviolet and photographed.

Sequencing analysis

To confirm that the fragments generated by PCR amplification with primers P3/P4 were mtDNA in origin, these bands were recovered from agarose gels and submitted to PCR reamplification with the original set of primers. Reamplification products were purified and sequenced using the ABI Prism BigDye Terminator Cycle Sequencing Kit (Perkin-Elmer, Foster City, CA) and an ABI Prism 377 DNA Sequencer (Perkin-Elmer). Sequencing was performed in both strands using the original primers.

Statistical analysis

Fisher’s exact test was used to assess the difference in different groups by the Scientific Package for Social Sciences (SPSS), version 10.0, Statistical analysis software. A value of P less than 0.05 was considered statistically significant.

Results

ΔmtDNA⁴⁹⁷⁷ deletions in lung cancer, adjacent normal and histologically normal lung tissue samples in subjects without lung cancer.

The detection of ΔmtDNA⁴⁹⁷⁷ between the origins of replication of light and heavy mtDNA strands was performed by PCR amplification with two sets of primers. To control the ability to PCR-amplify mtDNA, one primer pair (P1/P2) localized inside of the region referred to as ΔmtDNA⁴⁹⁷⁷ that amplified a 142-bp amplicon corresponding to wild-type mtDNA was used as an amplification control. In case the 4977-bp deletion is present, a 262-bp PCR product is generated as determined by another pair of primers (P3/P4) annealing to the fragments flanking the deleted region [Figure 1]. These bands were further sequenced and confirmed to be mitochondrial in origin. □mtDNA⁴⁹⁷⁷ deletions were detected in 54.1% (20/37) of lung cancers, 59.5% (22/37) of adjacent normal and 30.0% (6/30) of histologically normal lung tissues in subjects without lung cancer [Table 2]. No significant difference was found in the frequency of □mtDNA⁴⁹⁷⁷ deletions between the tumor and adjacent normal lung tissues (*P*

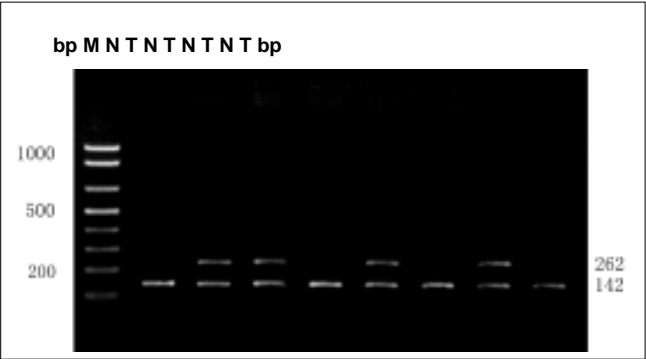


Figure 1: Detection of □mtDNA⁴⁹⁷⁷ in tumor and adjacent normal tissue

value = 0.815). The frequency of □mtDNA⁴⁹⁷⁷ deletions in lung cancers and adjacent normal tissues appeared to be higher than that in histologically normal lung tissue samples in subjects without lung cancer. However, because of limited number of samples, no significant difference was found between the tumor and normal lung tissues (*P* value = 0.101) and between adjacent normal and normal lung tissues (*P* value = 0.052).

The detection of □mtDNA⁴⁹⁷⁷ was performed by PCR amplification with two sets of primers. One primer pair (P1/P2) yields a 142 bp amplicon. In case the 4977 bp deletion is present, a 262-bp PCR product is generated as determined by another pair of primers (P3/P4). 142 and 262 bp bands, suggesting the presence of heteroplasmy, were amplified in 4977 bp deleted mtDNA and only 142 bp band was observed in wild mtDNA with primer sets P1/P2 and P3/P4. T: lung cancer tissues, N: matched normal tissues, M: PCR Markers.

Frequency of □mtDNA⁴⁹⁷⁷ and the age of patients

The frequency of □mtDNA⁴⁹⁷⁷ deletions increased with age in lung cancer, adjacent histologically normal and normal lung tissue samples and the highest frequency (88.9%) of it appeared in adjacent histologically normal tissue samples from the older patients (>65 years of age). The frequency of □mtDNA⁴⁹⁷⁷ deletions [Table 2] in tissue samples of the group over 55 years of age was

Table 2: The frequency of 4977 bp deletions and the age of patients													
Age	Lung cancer			Adjacent normal			Normal			Total			<i>P</i> value*
	+	-	+ (%)	+	-	+ (%)	+	-	+ (%)	+	-	+ (%)	
≥55	14	5	73.7	15	4	78.9	5	4	55.6	34	13	72.3	
<55	6	12	33.3	7	11	38.9	1	10	9.1	14	33	37.8	
Total	20	17	54.1	22	15	59.5	6	14	30.0	48	46		0.000

*obtained using the Fisher’s exact test, + □mtDNA⁴⁹⁷⁷ deletion, - without deletion, + (%) frequency of □mtDNA⁴⁹⁷⁷ deletion.

found to be significantly higher than that in the group younger than 55 years of age ($P < 0.001$).

Frequency of \square mtDNA⁴⁹⁷⁷ and cigarette smoke

The results suggested that cigarette smoke played an important role in the increase of \square mtDNA⁴⁹⁷⁷ in cancerous and normal tissues [Table 3]. The incidence of the \square mtDNA⁴⁹⁷⁷ in current smokers was significantly higher than in those of nonsmokers ($P = 0.002$). The incidence of the \square mtDNA⁴⁹⁷⁷ in ex-smokers (consisting of subjects who had been previously reported as smokers and had quit smoking for more than 1 year) was significantly higher than in those of nonsmokers ($P = 0.009$).

Frequency of \square mtDNA⁴⁹⁷⁷ deletions and histologically type of lung cancer

The \square mtDNA⁴⁹⁷⁷ deletions were detected in 16 of 23 squamous cell cancer and 4 of 14 adenocarcinoma samples [Table 4]. The incidence of the \square mtDNA⁴⁹⁷⁷ deletions in squamous cell cancers were significantly higher than in those of adenocarcinoma samples ($P = 0.0154$).

Discussion

The most common and also the most often assayed mtDNA deletion mutation, \square mtDNA⁴⁹⁷⁷ is a deletion that occurs between nucleotides 8 470 and 13 477 of the human mtDNA. It has been established as responsible for or associated with several human diseases, including ocular myopathy, Pearson's syndrome, diseases that progress with age.^[16] Slipped-strand mispairing (SSM), DNA damage and defective DNA repair are the causes producing \square mtDNA⁴⁹⁷⁷. The mispairing between two 13 bp direct repeats (positions 8 470-8 482 and 13 447-13 459) after a single-strand break caused by ROS or electron species produces fragment-deleted mtDNA.

The accumulation of somatic mtDNA mutations could contribute to the progression of mitochondrial diseases, the occurrence of various types of degenerative diseases and aging^[17-19] and could be

Table 4: The frequency of \square mtDNA⁴⁹⁷⁷ deletions and histologically type of lung cancer

Histologically type	+	-	+ (%)	<i>P</i> value*
Squamous cell cancer	16	7	69.5	0.0154
Adenocarcinoma	4	10	28.6	
Total	20	17		

*obtained using the Fisher's exact test, + \square mtDNA⁴⁹⁷⁷ deletion, - without deletion, + (%) frequency of \square mtDNA⁴⁹⁷⁷ deletion

associated with external factors such as radiation and cigarette smoking.^[20,21] Cortopassi *et al*^[22] reported that mtDNA 4977 deletions accumulate in normal individuals during aging, particularly in postmitotic tissues such as muscle and brain. The \square mtDNA⁴⁹⁷⁷ deletions in human lung tissues were found to be related to smoking habit or lifetime cigarette consumption.^[23] Cigarette smoke is a complex mixture of more than 3800 compounds, including both free radicals in high concentrations and chemical compounds that readily react to form other reactive substances.^[24] The free radicals cause peroxidation of membrane lipids, accumulation of oxidized dysfunctional proteins and increased DNA damage. In a study of the level of \square mtDNA⁴⁹⁷⁷ in bronchoalveolar tissues from smokers and non smokers a seven-fold higher frequency of this deletion was found in smokers.^[25] In this study, 37 matched lung cancer/adjacent histologically normal and 20 normal lung tissue samples from patients without lung cancer were analyzed by PCR technique and the results showed that the frequency of \square mtDNA⁴⁹⁷⁷ deletions is related to cigarette smoke and age. These data suggested that environmental and aging factors play an important role in the accumulation of mtDNA deletion mutations. The incidence of \square mtDNA⁴⁹⁷⁷ deletions in squamous cell cancers was higher than in those of adenocarcinoma samples. We suspected it may be just a reflection of the cigarette smoke influences operative during the accumulation of \square mtDNA⁴⁹⁷⁷, because the proportion (19/23) of smokers (including current smokers and ex-smokers) in patients with squamous cell cancer was higher than that (5/14) of adenocarcinoma.

Table 3: The frequency of \square mtDNA⁴⁹⁷⁷ deletions and cigarette smoke

	Lung			Normal			Total			<i>P</i> value*
	+	-	+ (%)	+	-	+ (%)	+	-	+ (%)	
Current smoker	11	4	73.3	4	5	44.4	15	9	62.5	0.999 ^a
Ex-smoker	6	3	66.7	2	2	50.0	8	5	61.5	0.009 ^b
Nonsmoker	3	10	23.1	0	7	0	3	17	15.0	0.002 ^c

*Obtained using the Fisher's exact test, + \square mtDNA⁴⁹⁷⁷ deletion, - without deletion, + (%) frequency of \square mtDNA⁴⁹⁷⁷ deletion. a: smoker comparative to Ex-smoker, b: Ex-smoker comparative to nonsmoker, c: Smoker comparative to nonsmoker.

Carcinogenesis is a multi-step process involving the accumulation of genetic changes that end in malignant cell transformation. Contribution of mtDNA mutations to carcinogenesis was postulated when wide spectra of the mtDNA alterations were reported in different types of cancers: colon, lung, pancreatic, liver, thyroid, bladder, prostate, esophageal and gastric cancer.^[7-11] Cavalli *et al*^[26] reported that the tumor cells were depleted of mitochondrial DNA by treatment with ethidium bromide. These rho(-) respiratory-deficient cells showed a distinct change in the tumorigenic phenotype, including loss of ability to grow in an anchorage-independent fashion and a substantial increase in sensitivity to cytotoxic drugs. Their results indicate that mitochondria/mitochondrial DNA play a direct role in modulating aspects of the tumorigenic phenotype. However, the biological impact of \square mtDNA⁴⁹⁷⁷ deletion on tumors is not entirely clear. Zhu *et al*^[27] found that the \square mtDNA⁴⁹⁷⁷ was present in 33% of adjacent histologically normal specimens from a cancerous breast and 46% of breast cancers. The 4977 bp mtDNA deletion was detected in 12 of 13 (92.3%) gastric tumor cell lines, 38 of 52 (73.1%) of gastric tumors and 27 of 52 (52%) adjacent normal tissues and was thought to play an important role in the carcinogenesis of human gastric tumor by Shen *et al*^[28] In this investigation, we found that \square mtDNA⁴⁹⁷⁷ was present in lung cancer, adjacent histologically normal and normal lung tissue samples and in eight cases of patients, the deletion was found in adjacent nontumoral tissues but not in cancerous tissues. These results suggest that \square mtDNA⁴⁹⁷⁷ deletion is not specific mutation to lung cancer.

The \square mtDNA⁴⁹⁷⁷ - which affects important genes involved in OXPHOS, such as ATPase 6, ATPase 8, cytochrome oxidase III, NADH subunits ND3, ND4, ND4L and ND5 and 5 of 22 tRNAs that are essential for protein synthesis of the mitochondria, may have a strong metabolic disadvantage so that cells carrying this mutation are selected against. Since each cell contains many mitochondria with multiple copies of mtDNA, it is possible that wild-type and mutant mtDNA can co-exist in a state called heteroplasmy. The mtDNA deletions accumulated in cells may result in impaired mitochondrial respiration and decreased ATP synthesis and the cells harboring high proportion of mtDNA deletions may not survive and lead to dropout from the population. Dani *et al* considered^[29] that though the metabolic effect of mtDNA 4977 deletion may be minimal in tissue, in tissue with active cell division, such as in tumors, even low levels of \square mtDNA⁴⁹⁷⁷ deletions may be intolerable. One could argue that tumor cells have a higher capacity for glycolysis and do

not rely entirely on OXPHOS to survive and hence that mutations such as \square mtDNA⁴⁹⁷⁷ deletion would not be metabolically detrimental. Nevertheless, the evidence^[30-32] indicates that, as glycolysis, OXPHOS is important to the survival and growth of tumors so that the cells harboring high proportion of mtDNA deletions cannot survive and lead to dropout from the population. “mtDNA4977 deletions were detected by Dani *et al*^[33] in 24% of the breast tumors, 52% of the colorectal tumors, 79% of the gastric tumors and 40% of the head and neck tumors as compared with 77, 83, 100 and 90% of the adjacent respective nontumoral tissues. Real-time Quantitative PCR experiments were further performed to quantify the number of “mtDNA4977 deletions per cell in selective nine cases of cancers, by determining the mitochondrial-to-nuclear DNA ratio. The average number (303.32) of deletions/cell of “mtDNA4977 in tumors was also found to be significantly lower than that (6.73) of the respective nontumoral tissue. Our data support a prior report^[34] indicating that the 4977 bp deletion is present at similar frequency in both normal and tumor tissue. Therefore, we conclude that \square mtDNA⁴⁹⁷⁷ deletion, which is not specific to lung cancer, may reflect the environmental and aging-process influences operative during lung cancer progression. As regards whether this deletion mutation is directly associated with the development and progression of lung cancer is still unclear and was not the purpose of our current study but is an area which we plan to investigate in the future.

References

1. Green DR, Reed JC. Mitochondria and apoptosis. *Science* 1998;281:1309-12
2. Tang L, Zhang Y. Mitochondria are the primary target in isothiocyanate-induced apoptosis in human bladder cancer cells. *Mol Cancer Ther* 2005;4:1250-9.
3. Howell N, Kubacka I, Mackey DA. How rapidly does the human mitochondrial genome evolve? *Am J Hum Genet* 1996;59:501-9.
4. Taylor RW, Turnbull DM. Mitochondrial DNA mutations in human disease. *Nat Rev Genet* 2005;6:389-402.
5. Parsons TJ, Muniec DS, Sullivan K, Woodyatt N, Alliston-Greiner R, Wilson MR, *et al*. A high observed substitution rate in the human mitochondrial DNA control region. *Nat Genet* 1997;15:363-8.
6. Cavalli LR, Liang BC. Mutagenesis, tumorigenicity and apoptosis: are the mitochondria involved? *Mutat Res* 1998;398:19-26.
7. Czarnecka AM, Golik P, Bartnik E. Mitochondrial DNA mutations in human neoplasia. *J Appl Genet* 2006;47:67-78.
8. Maitra A, Cohen Y, Gillespie SE, Mambo E, Fukushima N, Hoque MO, *et al*. The Human MitoChip: A high-throughput sequencing micro array for mitochondrial mutation detection. *Genome Res* 2004;14:812-9.
9. Frenny VJ, Antonella Z, Luisa A, Shah AD, Sheth JJ, Rocchi M. Cyto genetics and fluorescence in-situ hybridization in detection of hematological malignancies. *Indian J Cancer* 2003;40:135-9.
10. Dai JG, Min JX, Zhang GQ. The study on mitochondrial DNA

- Mutations in mouse tumors. *Zhong Hua Bing Li Xue Za Zhi* 2004;13:458-61.
11. Fliss MK, Usadel H, Cabellero OL. Facile detection of mitochondrial DNA mutations in tumors and bodily fluids. *Science* 2000;287:2017-9.
 12. Wu CW, Yin PH, Hung WY, Li AF, Li SH, Chi CW, *et al.* Mitochondrial DNA mutations and mitochondrial DNA depletion in gastric cancer. *Genes Chromosomes Cancer* 2005;44:19-28.
 13. Abnet CC, Huppi K, Carrera A, Armistead D, McKenney K, Hu N, *et al.* Control region mutations and the 'common deletion' are frequent in the mitochondrial DNA of patients with esophageal squamous cell carcinoma. *BMC Cancer* 2004;1:4-30.
 14. Maximo V, Soares P, Lima J, Cameselle-Teijeiro J, Sobrinho-Simoes M. Mitochondrial DNA somatic mutations (point mutations and large deletions) and mitochondrial DNA variants in human thyroid pathology: A study with emphasis on Hurthle cell tumors. *Am J Pathol* 2002;160:1857-65.
 15. Rogounovitch TI, Saenko VA, Shimizu-Yoshida Y, Abrosimov AY, Lushnikov EF, Roumiantsev PO, *et al.* Large Deletions in Mitochondrial DNA in Radiation-associated Human Thyroid Tumors. *Cancer Research* 2002;62:7031-41.
 16. Crott JW, Choi SW, Branda RF, Mason JB. Accumulation of mitochondrial DNA deletions is age, tissue and folate-dependent in rats. *Mutat Res* 2005;570:63-70.
 17. Linnane AW, Marzuki S, Ozawa T, Tanaka M. Mitochondrial DNA mutations as an important contributor to ageing and degenerative diseases. *Lancet* 1989;1:642-5.
 18. Wei Y H. Mitochondrial DNA alterations as ageing associated molecular events. *Mutat Res* 1992;275:145-55.
 19. Wei Y H. Oxidative stress and mitochondrial DNA mutations in human aging. *Proc Soc Exp Biol Med* 1998;217:53-63.
 20. Pang CY, Lee HC, Yang JH, Wei YH. Human skin mitochondrial DNA deletions associated with light exposure. *Arch Biochem Biophys* 1994;312:534-8.
 21. Lee HC, Lim ML, Lu CY, Liu VW, Fahn HJ, Zhang C, *et al.* Concurrent increase of oxidative DNA damage and lipid peroxidation together with mitochondrial DNA mutation in human lung tissues during aging-smoking enhances oxidative stress on the aged tissues. *Arch Biochem Biophys* 1999;362:309-16.
 22. Cortopassi GA, Arnheim N. Detection of a specific mitochondrial DNA deletion in tissues of older individuals. *Nucleic Acids Res* 1990;18:6927-33.
 23. Fahn HJ, Wang LS, Kao SH, Chang SC, Huang MH, Wei YH. Smoking-associated mitochondrial DNA mutations and lipid peroxidation in human lung tissues. *Am J Respir Cell Mol Biol* 1998;19:901-9.
 24. Pakhale SS, Jayant K, Bhide SV. Total particulate matter and nicotine in Indian bidis and cigarettes: a comparative study of standard machine estimates and exposure levels in smokers in Bombay. *Indian J Cancer* 1989;26:227-32.
 25. Ballinger SW, Boudier TG, Davis GS, Judice SA, Nicklas JA, Albertini RJ. Mitochondrial genome damage associated with cigarette smoking. *Cancer Res* 1996;56:5692-7.
 26. Cavalli, LR, Varella-Garcia M, Liang BC. Diminished tumorigenic phenotype after depletion of mitochondrial DNA. *Cell Growth Differ* 1997;8:1189-98.
 27. Zhu W, Qin W, Edward ER. Large-scale mitochondrial DNA deletion mutations and nuclear genome instability in human breast cancer. *Cancer Detect Prevent* 2004;28:119-26.
 28. Shen H, Zhao M, Dong B, Tang W, Xiao B, Liu JZ, *et al.* Frequent 4 977 bp deletion of mitochondrial DNA in tunlor cell lines, solid tumors and precancerous lesions of human stomach. *Zhonghua Yi Xue Za Zhi* 2003;83:1484-9.
 29. Dani SU, Dani MA, Simpson AJ. The common mitochondrial DNA deletion detamt DNA (4977) : Shedding new light to the concept of a tumor suppressor mutation. *Med Hypotheses* 2003;61:60-3.
 30. Hayashi JI, Takemitsu M, Nonaka I. Recovery of the missing tumorigenicity in mitochondrial DNA-less HeLa cells by introduction of mitochondrial DNA from normal human cells. *Somat Cell Mol Genet* 1992;18:123-9.
 31. Israel BA, Schaeffer WI. Cytoplasmic suppression of malignancy. *In Vitro Cell Dev Biol* 1987;23:627-32.
 32. Israel BA, Schaeffer WI. Cytoplasmic mediation of malignancy. *In Vitro Cell Dev Biol* 1988;24:487-97.
 33. Dani MA, Dani SU, Lima SP, Martinez A, Rossi BM, Soares F, *et al.* Less mtDNA4977 than normal in various types of tumors suggests that cancer cells are essentially free of this mutation. *Genet Mol Res* 2004;3:395-409.
 34. Bianchi MS, Bianchi NO, Bailliet G. Mitochondrial DNA mutations in normal and tumor tissues from breast cancer patients. *Cytogenet Cell Genet* 1995;71:99-103.