Study of desmoglein 1 and 3 antibody levels in relation to disease severity in Indian patients with pemphigus

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ABSTRACT

Objectives: To conduct a cross-sectional study to compare Dsg1 and Dsg3 antibody levels independently with severity of disease activity in pemphigus vulgaris (PV) and pemphigus foliaceus (PF). Methods: Blood samples from 44 patients with pemphigus (PV-38, PF-6) were analyzed using ELISA. The severity of skin and mucosal disease was graded using a score from 0 to 3. Results: A statistically significant correlation between increase in Dsg 3 antibody titres with severity of oral involvement and Dsg 1 titres with severity of skin involvement was found in both PV and PF patients (p<0.01). However, we were unable to demonstrate a relationship between increased titres of Dsg1 and Dsg 3 antibodies with oral and skin involvement respectively. Conclusion: This study suggests that the severity of skin and oral disease in pemphigus is determined by the quantities of Dsg1 and Dsg3 antibodies respectively.

Key Words: Antibodies, Desmoglein 1 and 3, Pemphigus, Serology

INTRODUCTION

Pemphigus is one of the best-characterized tissue-specific autoimmune diseases. The desmoglein proteins desmoglein-3 (Dsg3) and desmoglein-1 (Dsg1) have been identified as the antigens in pemphigus vulgaris (PV) and pemphigus foliaceus (PF) respectively. While earlier studies suggested that indirect immunofluorescence titers were a useful marker for disease activity in pemphigus,[1-3] a detailed study concluded that serial titers were not consistent enough to be used as a guide for therapy and prognosis.[4] The production of recombinant Dsg1 and Dsg3 molecules has provided the opportunity to determine levels of antibodies to them (anti-Dsg1 IgG and anti-Dsg3 IgG respectively) and to see if they correlate with disease severity.[5] However, there are conflicting data in this regard in many studies using immunofluorescence titers.[1-3]

ELISA is a simple and effective tool for the quantitative analysis of antibody levels. Harman et al. used ELISA to measure serial Dsg1 and Dsg3 antibody levels and correlate them with the severity of oral and skin ulceration in PV and PF.[6] As there are hardly any similar studies from India, we conducted this cross-sectional study to correlate the severity of oral and/or cutaneous involvement in patients with PV and PF with ELISA levels.
values for antibodies to Dsg1 and Dsg3 independently.

**METHODS**

Forty-four patients with pemphigus (38 with PV and 6 with PF, diagnosed based on clinical examination, histopathology and direct immunofluorescence) and three patients with extensive burns (as controls) were studied. We chose patients with burns as controls because pemphigus-like circulating intercellular antibodies have been found in such patients. ELISA was used to confirm the presence of Dsg1 and Dsg3 autoantibodies in PF and PV patients and an attempt was made to correlate their levels with cutaneous or mucosal involvement and disease activity. The blood samples were taken from patients who were untreated and with active disease or from those with resolving disease on treatment of varying durations. No serial samples were taken from any patient.

A simple arbitrary scoring system was used to grade the severity of skin or mucosal involvement at the time of sampling as follows:

**Oral score**
0 = No mucosal involvement.
1 = Minimal disease (only buccal mucosal, labiogingival, lingual, palatal or pharyngeal involvement).
2 = Moderate disease (buccal and labiogingival, lingual, palatal or pharyngeal involvement).
3 = Severe disease (extensive oral erosions, i.e., >3 mucosal sites affected)

**Skin score**
0 = Quiescent disease.
1 = Minimal disease (≤10% body surface area involvement (BSA) involved).
2 = Moderate disease (11-30% BSA involved).
3 = Severe disease: (>30% BSA involved).

Antibodies to Dsg1 and Dsg3 were detected in the sera of patients by the micro ELISA technique using kits from Medical and Biological Laboratories Co. Ltd., Japan, as per the manufacturer’s instructions. ODs were adjusted relative to the reference sera supplied with the kits.

Forty-seven serum samples were analyzed. The serum samples were randomly taken and subsequently stored at -20°C before processing them for ELISA.

The unpaired t test was used to compare Dsg1 and Dsg3 levels in relation to severity of either skin or mucosal lesions.

**RESULTS**

A cut off value for each ELISA was established from the mean ± 2SD of the three patients with extensive burns. Although the cut off values given in the kit were taken as standard, to increase the sensitivity, we wanted to generate a cut off value from the local population as the kit was manufactured elsewhere. For Dsg1 ELISA, the mean was 0.143 and for Dsg3 it was 0.102. This resulted in a cut off value of 0.406 for Dsg1 ELISA and 0.212 for Dsg3 ELISA. Dsg1 and Dsg3 antibodies were found to be present in all the patients of PV and PF. A summary of the disease and demographic parameters is given in Table 1.

For Dsg1 ELISA, 5 of 6 (83.3%) PF patients and 23 of 38 (60.52%) PV patients were above the cut off values. For Dsg3 ELISA, 31 of 38 (81.57%) PV patients and 1 of 6 (16.6%) PF patients exceeded the cut off value.

**ELISA values and disease severity**

We attempted to correlate the extent of skin and mucosal involvement [Table 2] with the independent values of Dsg1 and Dsg3 ELISA [Table 3]. Although a wide range of values was obtained, the general trend was that Dsg3 levels were higher in patients with extensive oral mucosal involvement, i.e., patients with moderate and severe mucosal involvement had

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**Table 1: Disease and demographic parameters**

<table>
<thead>
<tr>
<th>Pemphigus vulgaris (n=38)</th>
<th>Pemphigus foliaceus (n=6)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Males</td>
<td>18</td>
</tr>
<tr>
<td>Females</td>
<td>20</td>
</tr>
<tr>
<td>Age (years)</td>
<td>10-52</td>
</tr>
<tr>
<td>Active disease (untreated)</td>
<td>26</td>
</tr>
<tr>
<td>Healing/inactive disease</td>
<td>12</td>
</tr>
</tbody>
</table>
significant higher Dsg3 levels than those with mild or no mucosal involvement (p<0.01). However, Dsg1 ELISA levels did not always correlate with the severity of mucosal involvement (p>0.1) [Table 2].

Analysis showed a direct relationship between the severity of skin involvement and the levels of the Dsg1 antibodies, i.e., patients with moderate and severe disease activity had significant higher levels of Dsg1 antibodies when compared to those with minimal disease activity (p<0.01). However, ELISA levels of antibodies to Dsg3 did not show a consistent relation to the extent of skin involvement (p>0.1), although patients with extensive disease had slightly more elevated levels of antibodies to Dsg3 [Table 3].

After analyzing the overall relation between the desmoglein antibody levels and disease severity in terms of skin or mucosal involvement, the data was also analyzed to determine whether the relationship between the levels of desmoglein antibodies and disease severity was dependent on the type of pemphigus. The difference between PF and PV was not significant when the severity of skin involvement and Dsg-1 antibody levels were correlated (p>0.1).

However, the presence of severe mucosal involvement correlated well with increased Dsg3 antibody values in PV patients (p<0.05). All six of our patients with PF (none with mucosal involvement) had detectable levels of Dsg3 antibodies.

**DISCUSSION**

The autoimmune target of pemphigus is desmoglein, a cadherin type of cell-to-cell adhesion molecule found in the desmosomes. There are three isotypes of desmoglein: Dsg1, Dsg2 and Dsg3. Dsg1 and Dsg3 are usually restricted to the stratified squamous epithelia, where blister formation is located in pemphigus.[7] Patients with pemphigus have anti-Dsg1 and/or anti-Dsg3 IgG autoantibodies. IgG1 and IgG4 are the most common subclasses of antibodies in patients with active disease, but IgG4 is more pathogenic and indicates the activity of the disease. Even though PV and PF are distinct diseases, there are several case reports that suggest that a shift between PF and PV may occur. In some of these reports, immunoblotting studies suggested that the antigen has changed in accordance with the change in clinical picture.

Our data demonstrates that in pemphigus, skin disease severity relates to Dsg1 levels, irrespective of the type of pemphigus (PF/PV). Moreover, Dsg3 levels correlate with mucosal severity in patients with PV. Our findings are in consonance with the study by Harman et al.[6] who also found that levels of antibodies to Dsg1 correlated with an increase in the severity of skin involvement and elevated anti-Dsg3 antibodies to the severity of mucosal involvement.

Recently, according to the desmoglein compensation theory, pemphigus has been divided into mucosal predominant PV, in which autoantibodies are present only to Dsg3; mucocutaneous PV, where antibodies to both Dsg1 and Dsg3 are present; and PF, where only antibodies to Dsg1 are present and not to Dsg3.[8-10]

Correlation of the severity of disease with the level of antibodies detected in patients with PV resulted in two interesting observations: Nine out of the 12 patients with PV having quiescent and resolving disease still had high ELISA values for Dsg3 antibodies, although they had been free from active disease for approximately 6 months or more. The
reasons for this could be, firstly, the polyclonal nature of the pemphigus autoantibody; therefore, the IgG subclass may be an important determinant. Moreover, up to 70% of the first degree relatives of patients with pemphigus have detectable antibodies that are not of the IgG4 subtype.\[11\] Secondly, as stated by Harman et al.,\[6\] treatment may have dampened pathogenic mechanisms triggered by antibody binding. Another interesting observation was that all our 6 PF patients, both with active and quiescent disease, demonstrated the presence of autoantibodies to Dsg3. Arteaga et al. demonstrated autoantibodies to Dsg3 in 19 of the 276 patients of PF and fogo selvagem reviewed by them.\[12\] This suggests that two separate species of antibodies are present rather than one antibody, capable of cross-reacting with Dsg1 and Dsg3. Moreover, their study showed that affinity of purified antibodies to Dsg3 from patients with PF and fogo selvagem by passive transfer induced a PV-like disease in mice. These results suggest that a subset of patients with PF have antibodies to Dsg3 that may also be involved in the pathogenesis of cutaneous disease. Another explanation for this could be that probably there is an epitope shift from PF to PV occurring in these patients. It will be interesting to see if these subsets of patients show a change in their clinical presentation on long-term follow-up (epitope shift phenomenon). Long-term studies are required to confirm these findings.

In a few cases of PV with active disease, low ELISA values for Dsg1 and Dsg3 were found; it is possible that the sera contained pathogenic antibodies to non-desmoglein molecules or to intracellular domain of Dsg1/Dsg3, which could have been undetectable by the ELISA used to detect specific antibodies to only Dsg 1 and Dsg 3.

Our findings suggest that at any given period, the level of the antibodies does correlate with disease severity. However, this may not be true for a small proportion of cases because of the reasons explained above. A detailed prospective study for evaluating the ELISA levels of a given patient for prolonged periods would be helpful in actual correlation with the disease severity and changes that may occur in the spectrum of the disease.

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REFERENCES