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Estimation of phylogenetic relationships of *Phlomis* species based on seed protein polymorphism

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Abbreviations: DTT: dithiothreitol

RAPDs: randomly amplified polymorphic DNA SDS-PAGE: sodium dodecyl sulphate polyacrylamide gel electrophoresis TES: TE-saline UPGMA: unweighted pair-group method with arithmetic averages

In this study, phylogenetic relationships among 39 Phlomis taxa were investigated based on seed protein profiles produced by sodium dodecyl sulphate polyacrylamide gel electrophoresis (SDS-PAGE). A total of 21 polypeptide bands were scored, of which, 19 were polymorphic among the taxa of the genus Phlomis. A distance matrix was generated from the similarity matrix which was computed by using Jaccard's similarity coefficients, based on polymorphic bands and then an UPGMA tree was established through cluster analysis performed on the distance matrix. Genetic distances ranged from 0.00 to 0.50 within subsection Dendrophlomis; from 0.00 to 0.625 within subsection Gymnophlomis and from 0.00 to 0.769 within subsection Oxyphlomis. The UPGMA tree formed four groups. The topology of the tree is in agreement with the taxonomic view regarding the section Phlomis as it is divided into three subsections as Dendrophlomis, Gymnophlomis and Oxyphlomis based on morphological characters. The grouping pattern of the tree also indicated that subsection Dendrophlomis is more closely related to subsection Gymnophlomis than subsection Oxyphlomis.

The genus Phlomis L. contains over 100 species that have been divided into two main sections: Phlomoides and Phlomis (Moench, 1794; Albaladejo et al. 2005). Section Phlomis was further subdivided into three subsections, Dendrophlomis, Gymnophlomis and Oxyphlomis (Bentham, 1834). The diagnostic character for separating sections is corolla shape. Species of section Phlomis, which have corolla with curved upper lip and trifid lower lip with large median and smaller lateral lobes, differs from species of section Phlomoides that have corolla with straight upper lip and trifid lower lip with sub equal lobes (Azizian and Moore, 1982). Section Phlomis occurs mostly in the Mediterranean region, where Turkey is regarded as one of the primary center of diversification for species of the section (Hedge, 1986). For instance, in the Flora of Turkey, the genus Phlomis consists of 34 species and six varieties, of which 22 are endemic to that flora (Huber-Morath, 1982).

Taxonomic relationships among species of the genus *Phlomis* have been assessed by morphological characters (Azizian and Moore, 1982; Huber-Morath, 1982; Taylor,

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Figure 1. Seed protein profiles of 39 *Phlomis* taxa (M = size marker; numbers 1-22 as in Table 1).

1998). Nevertheless, based on morphological characters alone, it is difficult to distinguish P. armeniaca, P. sieheana, P. sintenisii, P. lanceolata and P. carica from each other within subsection Gymnophlomis because they have overlapping variations in terms of the major delimiting morphological characters such as plant height, basal leaves sizes and shapes, bracteol length, calyx teeth shape and length, indumentum of calyx and number of flowers per verticillaster (Huber-Morath, 1982). Morever, differentiation of P. carica from P. armeniaca, and of P. nissoli from P. syriaca within this complex is based on indimentum of calvces and leaves; hovewer, this character shows high variation among individuals of a population and among populations of these species (Huber-Morath, 1982). Also, hybridization is a common incident in this genus. leading to the formation of many interspecific hybrids that increase the taxonomic complexity (Albaladejo et al. 2004; Albaladejo et al. 2005; Albaladejo and Aparicio, 2007; Yuzbasioglu et al. 2008b). Seed storage protein markers have been sucessfully used to resolve taxonomic relationships and characterize cultivated varieties in a number of crop plant species (Igrejas et al. 1999; Vladova et al. 2000; Jha and Ohri, 2002; Karihaloo et al. 2002; Syros et al. 2003; Bhargava et al. 2005; Cherdouh et al. 2005; Alvarez et al. 2006; Stoilova et al. 2006; Mirali et al. 2007; Rout and Chrungoo, 2007; Yuzbasioglu et al. 2008a) because they are stable, uniform, reliable, reproducible and largely independent of environmental fluctuations (Ghafoor et al. 2002; Panigrahi et al. 2007). The taxonomic relationships among species of Phlomis based on seed protein electrophoresis have not been studied so far. The objective of the present study was to assess taxonomical relationships among 39 Phlomis taxa based on sodium dodecyl sulphate polyacrylamide gel electrophoresis (SDS-PAGE) of seed proteins.

MATERIALS AND METHODS

Seeds from 39 *Phlomis* taxa examined in the present study were collected from different parts of Turkey (Table 1). Seeds from each taxon were collected from at least 10 single plants within a population of that taxon and then the seeds of each taxon were bulked. The bulked seeds of each

taxon were ground into a fine powder with liquid nitrogen. 25 mg of flour samples were transferred into 200 µL of TE-Saline (TES) buffer within 1.5 ml eppendorf tubes containing 100 mM Tris HCl, 50 mM NaCl, 1 mM EDTA, 2% SDS, 2% PVP, 100 μM Nα-P-Tosyl-L-Lysine Chloromethyl Ketone (TLCK), 0.3 μM Phenylmethanesulphonyl Fluoride (PMSF) and 100 mM Phenylalanine Ketone (TPCK). The flour samples were mixed with the TES buffer by vortexing for 10 min at room temperature. Then the mixture was heated for 3 min at between 80 and 90°C. After cooling at room temperature, 4 μ L of DTT (dithiothreitol) solution (3.25 M DTT in dH₂O) was added to the mixture. Having centrifugated the tubes at 15000 x g for 15 min at 4°C by using a 3K30 cooling centrifuge, the spernatant was transferred into new tubes. Then protein content of each sample was determined according to the modified Bradford assays described by Ramagli and Rodriguez (1985) using Bovine Serum Albumin (BSA) faction as the standard. The protein concentration of each sample was equalized by TES buffer and the equalized protein concentration was either mixed in half with loading buffer (65 mM Tris HCl, pH 6.8, 1% SDS, 2% 2-mercaptoethanol, 20% Glycerol) or kept on ice and stored at -20°C for further use.

Protein patterns were analyzed by 14% SDS-PAGE (18 x 20 x 0.1 cm; vertical slab gels; Protean Xii, BioRad, CA, USA) according to Laemmli (1970). The molecular weights of the dissociated polypeptides were determined by using molecular size markers in the range between 14.4 and 116 kD (Fermentas SMO431). Gels were stained by a sensitive colloidal Coomassie Brillant Blue G-250 as described by Olivieri et al. (2001) and photographed using a digital image analysis system (Vilber-Lourmat, Bio-Gene V.99). A similarity matrix based on Jaccard's similarity coefficient was produced with NTSYS-pc (Rohlf, 1992) from protein bands scored as 0 (absent) and 1 (present) and then, a distance matrix was constructed by subtracting the similarity coefficients from 1. A dendrogram based on the distance matrix was produced using the unweighted pair-





Figure 2. Seed protein profiles of 39 *Phlomis* taxa (M = size marker; numbers 23-39 as in Table 1).

Table 1. *Phlomis* material and its source.

Numbers	Collector Number	<i>Phlomis</i> taxa	Collection site	Altitude (m)	Date				
1	MYD 1718	P. russeliana (Sims) Bentham	Bolu; Around Abant Lake.	1220	12 viii 1999				
2	MYD 1451	P. fruticosa L.	Izmir; Cesme, Ciftlikkoy.	60-70	27 iv 2000				
3	MYD 1102	<i>P. lunariifolia</i> Sm.	Antalya; Alanya, East slopes of Kaledran stream.	50	25 vi 1998				
4	MYD 1166	<i>P. grandiflora</i> var. <i>grandiflora</i> H.S.Thompson	Antalya; Kas-Elmali road. Oluk Yaylasi, between Gomuce and Kuruova villages.	1700	28 vi 1998				
5	MYD 1103	P. grandiflora var. fimbrilligera HubMor.	Antalya; Alanya, West slopes of Kaledran stream.	60	25 vi 1998				
6	MYD 1190	<i>P. viscosa</i> Poiret	Osmaniye; Osmaniye-Yarpuz Yaylasi road.	550	6 viii 1998				
7	MYD 1678	P. bourgaei Boiss.	Antalya; Kemer, around Agva stream.	60	4 vii 2003				
8	MYD 1100	P. leucophracta P.H. Davis & HubMor.	Antalya; Between Anamur-Alanya, around Tenzile village.	150	25 vi 1998				
9	MYD 1065	P.longifolia var. longifolia Boiss. & Bl.	Hatay; Between Gedik and Atik.	800	13 vi 1998				
10	MYD 1075	<i>P.longifolia</i> var. <i>bailanica</i> (Vierh.) Hub Mor.	Hatay; Above Guzelyayla (Sogukoluk).	980	15 vi 1998				
11	MYD 1158	<i>P. amanica</i> Vierh.	Hatay; Arsuz (Ulucinar), Haymaseki village, around Aktepe.	250-300	9 viii 1998				
12	HD 6253	<i>P. lycia</i> D. Don	Antalya; Termessos National Park.	300-400	22 vii 1996				
13	MYD 1096	P. monocephala P.H. Davis	İçel; Between Kazanci-Anamur, around Abanoz Yaylasi.	1400	25 vi 1998				
14	HD 6643	P. chimerae Boissieu	Antalya; Kemer, around Cirali.	20-50	15 v 1998				
15	MYD 1029	P. oppositiflora Boiss. & Haussk.	Sivas; Between Darende-Gurun.	1390	8 viii 1997				
16	MYD 1287	P. bruguieri Desf.	Gaziantep; Soft Dagi.	1350- 1460	16 vi 1999				
17	MYD 1349	<i>P. armeniaca</i> Willd.	Tokat; Turhal, East slopes of Hastahane hill.	650	3 vii 1999				
18	MYD 1384	P. physocalyxHubMor.	Sivas; Gurun-Kayseri 23 km.	1600	11 vii 1999				
19	MYD 1251	P. angustissima HubMor.	Afyon; Dirmil; Dirmil pass.	1580	9 vi 1999				
20	MYD 1716	P. capitata Boiss.	Malatya; Venk village.	1195	12 vii 2003				
21	MYD 1178	P. kotschyana HubMor.	Hatay; Arsuz, Haymaseki village, around Aktepe.	250-300	19 v 1999				
22	MYD 1463	P. sieheana Rech.	Kayseri; Gesi, around Ildem Koop.	c. 1150	28 vi 2000				
23	MYD 1405	P. sintenisii Rech.	Elazig; Around Harput Castle.	1500	12 vii 1999				
24	MYD 1436	P. lanceolata Boiss. & Hohen.	Van; Van-Gurpinar 11 km, Karabas	2225	20 vii 1999				

25	MYD 1352	<i>P. linearis</i> Boiss. & Bal.	Kayseri; Develi-Kayseri 12 km (Erciyes road).	1930	9 vii 1999
26	MYD 1296	P. brunneogaleata HubMor.	Gaziantep; Soft Dagi.	1350- 1460	16 vi 1999
27	MYD 1086	P. nissolii L.	Nigde; Bor-Aksaray road, between Yesilyurt- Altunhisar villages.	1100	24 vi 1998
28	MYD 1063	P. syriaca Boiss.	Adana; Ceyhan, Yilankale.	100-300	12 vi 1998
29	MYD 1036	<i>P. kurdica</i> Rech.	Kahramanmaras; Elbistan-Goksun 13 km (old road).	1140	7 viii 1997
30	MYD 1125	<i>P. carica</i> Rech.	Denizli; Acipayam, Alaattin town, Degirmendere site.	1100	28 vi 1998
31	MYD 1055	P. tuberosa L.	Sivas; Imranlı-Zara 10 km.	1350	16 viii 1997
32	MYD 1163	<i>P. samia</i> L.	Adana; Gulek village-Adana 1 km.	1100	5 vii 1998
33	MYD 1321	P. punges var. pungens Willd.	Kayseri; Yahyali-Maden road 14 km.	1490	27 vi 1999
34	MYD 1057	P. punges var. laxiflora Velen.	B4 Ankara; Hacettepe university Beytepe campus.	1050- 1200	19 viii 1997
35	MYD 1034	P. punges var. hirta Velen.	Kahramanmaras; Cardak-Elbistan 5 km.	1240	7 viii 1997
36	MYD 1320	P. punges var. hispida HubMor.	Kayseri; Yahyali-Maden road 14 km.	1490	27 vi 1999
37	MYD 1429	P. punges var. seticalycina Nab.	Van; Gurpinar-Catak 25 km.	2090	20 vii 1999
38	MYD 1025	<i>P. integrifolia</i> HubMor.	Malatya; Malatya-Arapkir 30 km, around Kuruçay.	770	7 viii 1997
39	MYD 1038	P. rigida Labill.	Malatya; Elbistan-Malatya 53 km.	1600	7 viii 1997

group method with arithmetic averages (UPGMA) under the NJ subprogram in the PHYLIP software package version 3.6a3 (Felsenstein, 2002).

RESULTS AND DISCUSSION

Seed storage proteins of the 39 Phlomis taxa examined by SDS-PAGE in the present study produced reproducible and stable protein bands that were used for the classification of the taxa studied in the genus (Figure 1 and Figure 2). A total of 21 polypeptide bands were scored, of which, 2 were universally present and 19 were polymorphic. The cluster analysis of the taxa based on distance matrix produced from Jaccard's coefficient was represented by an UPGMA dendrogram in Figure 3. The dendrogram formed four groups. The first group consisted of taxa from section Phlomis subsection Dendrophlomis: P. russeliana, P. lycia, P. fruticosa, P. grandiflora var. grandiflora, P. bourgaei, P. chimerae, P. lunariifolia, P. viscosa, P. grandiflora var. fimbrilligera, P. longifolia var. longifolia, P. longifolia var. bailanica, P. amanica, P. monocephala and P. leucophracta. Except for P. russeliana, a perennial herb, all other taxa within this subsection are shrubs. In species of this subsection, verticillasters generally have 12-20 flowers and the number of verticillasters range between 1 and 3. In species of this subsection, bracteoles are linear-subulate to lanceolate and ovate in shape, generally 10-18 mm in length and the number of bracteoles is generally between 15 and 30. Relationships among P. viscosa, P. lunariifolia. P. grandiflora var. fimbrilligera, P. longifolia var. longifolia, P. amanica and P. monocephala obtained by SDS-PAGE analysis in the present study were generally in agreement with the previous study of relationships among species of *Dendrophlomis*, based on randomly amplified polymorphic DNA (RAPDs) markers (Yuzbasioglu and Dadandi, 2008). Nevertheless, different groupings were observed among P. bourgaei, P. leucophracta, P. chimerae and P. lycia when compared with the findings of the previous study, which excluded samples of P. russeliana, P. fruticosa and P. longifolia var. bailanica. Based on previous RAPD analysis, the grouping of these four taxa agrees better with the traditional taxonomic classification than grouping them based on SDS-PAGE analysis. SDS-PAGE clustering of the species of subsection Dendrophlomis into two groups is consistent with the groupings of the previous RAPD study. However, P. leucophracta and P. chimerae were not clustered in the same group. In contrast, the finding from previous RAPD

Table 2. Genetic distances	(as percentages) among 39 <i>Phlomis</i> taxa in th	e genus <i>Phlomis</i> (T	The numbers 1-39 refer to Table	ə 1)
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Γ	1	2	3	4	5	6	7	В	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37	38	39
1	-			Γ																					Γ												\square	\square	
2	22	F																									\square										\square	\square	
β	40	40	F	Γ	Γ								\square		\square				Γ		Γ	Γ	Γ	Γ	Γ	Γ	Γ										\square	\square	Γ
4	13	13	33	F									\square														\square												
Б	33	33	13	25	F								\square						Γ						Γ		\square										\square	\square	
6	22	22	22	13	13	F																																	
7	33	13	50	25	44	33	F						\square		\square				Γ				Γ	Γ	Γ	Γ	Γ										\square	\square	
β	38	38	38	29	29	38	29	F	Γ	Γ	Γ	Γ	Γ		Γ		Γ	Γ	Γ	Γ	Γ	Γ	Γ	Γ	Γ	Γ	Γ						Γ				\square	Γ	Γ
9	33	33	13	25	25	33	44	29	F																														
10	<u> 3</u> 3	33	13	25	25	33	44	29	þ	F																													
11	33	33	13	25	25	33	44	29	þ	þ	-																												
12	211	11	45	22	40	30	22	44	40	40	40	-	\square														\square												
13	325	25	25	14	14	25	38	17	14	14	14	<u>β</u> 3	F												\square														
14	40	22	40	<u>β</u> 3	33	40	33	38	<u>β</u> 3	33	33	30	25	-																									
15	560	50	64	60	60	64	60	67	60	60	60	40	56	50	-																								
18	65	65	65	64	60	65	64	70	64	64	64	45	60	55	13	-											\square												
17	60	50	64	60	60	64	60	67	60	60	60	40	56	50	ρ	13	-								\square														
18	866	56	70	67	67	70	50	67	67	67	67	44	63	56	29	38	29	-	\square			\square	\square	\square	\square	\square	\square		\square	\square									
19	966	38	56	50	50	56	50	67	60	50	60	44	43	38	29	38	29	33	F																				
20	173	60	60	70	70	73	66	63	66	66	66	64	67	60	56	60	66	43	43	-																			
21	50	50	64	60	60	64	60	67	60	60	60	40	56	33	25	33	25	29	29	56	-				\square		\square												
22	2 50	50	64	60	60	64	60	67	60	60	60	40	56	33	25	33	25	29	29	56	p	F			\square														
23	344	44	60	66	56	60	66	63	66	56	66	<u>β</u> 3	50	44	14	25	14	17	17	50	14	14	-	\square	\square	\square	\square												
24	44	44	60	66	56	60	66	63	66	56	66	<u>β</u> 3	50	44	14	25	14	17	17	60	14	14	p	F			\square												
25	560	60	60	60	44	60	60	67	60	60	60	40	56	50	25	13	25	29	29	66	25	25	14	14	F		\square												
28	64	50	50	60	60	64	60	67	44	44	44	65	56	50	44	50	44	50	29	14	44	44	38	38	44	F													
27	44	44	60	66	56	60	66	63	66	56	66	<u>β</u> 3	50	44	14	25	14	17	17	50	14	14	p	p	14	38	F		\square	\square									
28	3/50	50	50	60	44	60	60	67	60	60	60	40	56	50	25	13	25	29	29	56	25	25	14	14	p	44	14	-											
29	960	60	25	66	38	44	70	63	<u>β</u> 8	38	38	64	50	60	56	44	66	63	43	50	66	56	60	60	38	38	60	38	-										
30	64	50	50	60	60	64	60	67	44	44	44	65	56	50	25	33	25	50	29	38	44	44	38	38	44	25	38	44	38	-									
31	70	82	70	80	67	70	80	75	80	80	80	73	78	82	67	56	67	67	75	78	67	67	63	63	60	80	63	50	63	80	-								
32	282	82	56	80	67	70	80	75	67	67	67	83	78	82	80	70	80	75	75	63	80	80	78	78	67	67	78	67	43	67	67	F							
33	373	73	81	80	80	81	71	77	80	80	80	67	79	73	50	54	50	65	67	69	62	62	58	58	62	71	68	62	79	62	67	77	ŀ						
34	173	73	81	80	80	81	71	77	80	80	80	67	79	73	50	54	50	65	67	69	62	62	58	68	62	71	68	62	79	62	67	77	þ	-					
35	571	71	80	79	79	80	69	75	79	79	79	64	77	71	45	50	45	50	64	67	68	68	65	65	68	69	65	58	77	58	64	75	9	9	-				
38	671	80	80	79	79	80	79	75	79	79	79	73	77	80	58	62	58	64	75	77	69	69	67	67	69	79	67	69	77	69	64	75	9	9	18	F			
37	71	80	80	79	79	80	79	75	79	79	79	73	77	80	58	62	58	64	75	77	69	69	67	67	69	79	67	69	77	69	64	75	9	9	18	0	-		
38	375	85	85	83	83	85	83	80	83	83	83	77	82	85	60	64	60	67	80	82	73	73	70	70	73	83	70	73	82	73	50	67	36	36	30	30	30	ŀ	
39	967	67	67	73	64	67	73	79	73	73	73	60	71	67	42	33	42	68	58	71	54	54	50	50	42	64	50	42	62	54	68	69	23	þ3	17	β1	β1	42	\mathbf{F}

study placed both *P. leucophracta* and *P. chimerae* in one group and the rest of the species of *Dendrophlomis* into another group. *P. longifolia* var. *longifolia*, *P. longifolia* var. *bailanica* and *P. amanica* displayed the same protein pattern based on genetic distances obtained from SDS-PAGE analysis. Morphologically, they are similar to each other in terms of the absence of glandular hairs. The highest genetic distance within subsection *Dendrophlomis* was observed between *P. bourgaei* and *P. lunariifolia* with a genetic distance value of 0.50 in the present analysis (Table 2). *P. bourgaei* has glandular hairs especially on the floral region (calyx and bracteoles) while *P. lunariifolia* is eglandular. Bracteoles are 10-24 mm in length and 0.75-1.5 mm in width in *P. bourgaei* and 7-15 mm in length and 2-

3.5 mm in width in *P. lunariifolia*. Calyx teeth lengths are between 3-9 mm in *P. bourgaei* and 2-5 mm in *P. lunariifolia*. The range of genetic distances computed from the present study is concordant with the range (0.145-0.495) obtained from our previous RAPD analysis, however, the lowest genetic distances found between *P. amanica* and *P. monocephala* and the highest between *P. chimerae* and *P. lunariifolia* in the previous study. Differences observed in the groupings of the taxa of subsection *Dendrophlomis* in the dendrograms obtained from SDS-PAGE analysis and the previous RAPD study could be caused by several reasons. For instance, seed proteins are coded by several homologous multigene families (Panigrahi et al. 2007), but RAPD bands may



Figure 3. UPGMA dendrogram of 39 *Phlomis* taxa based on distances produced from Jaccard' similarity coefficients of seed storage proteins.

cover coding and non coding regions of DNA (Kirsten et al. 1998; Bussell et al. 2005). In addition to this, the different accessions sampled in the two studies, the different methods used for estimating genetic distances and the number of loci sampled in the two studies could be some other reasons of these differences.

The second group in the dendrogram comprised taxa from section **Phlomis** subsection Gymnophlomis: Ρ. oppositiflora, P. armeniaca, P. bruguieri, P. kotschyana, P. sieheana, P. sintenisii, P. lanceolata, P. nissoli, P. linearis, P. syriaca, P. physocalyx, P. angustissima, P. capitata, P. brunneogaleata, P. carica and P. kurdica. Species in section Phlomis subsection Gymnophlomis are perennial herbs. In species of this subsection, bracteoles are weak and subulate to linear-lanceolate in shape, 6-12 in number and 0.5-10 mm in length, but they may be absent, as in P. oppositiflora or may reach up to 25 mm, as in P. bruguieri (Azizian and Moore, 1982; Huber-Morath, 1982). Verticillasters are usually 4-8 flowered in these species and the number of verticillasters per stem is between 3 and 6 (Huber-Morath, 1982).

In the present study, no variability was observed between *P. kotschyana* and *P. sieheana*, between *P. linearis* and *P. syriaca* and among *P. sintenisii*, *P. lanceolata* and *P. nissoli* within subsection *Gymnophlomis*. In *P. kotchyana* and *P. sieheana*, bracteoles are 2-3 mm in length and leaves length is three times longer than their width in both species. On the other hand, the base of the basal and cauline leaves is cordate in *P. kotschyana* but cuneate or truncate in *P.*

sieheana. The shapes of basal and cauline leaves are cordate to broadly triangular cordate in P. kotschyana but oblong to oblong-lanceolate in P. sieheana. In P. sintenisii, P. lanceolata and P. nissoli bracteoles are stellatetomentose and these species are also eglandular. Basal leaves are oblong to oblong-lanceolate in P. sintenisii and P. lanceolata but oblong to ovate in P. nissoli. Calyx teeth are lanceolate-acuminate in P. sintenisii and P. lanceolata but it is ovate-triangular in P. nissoli. P. linearis and P. syriaca are eglandular; their bracteoles are stellatetomentose and their calyx is slightly campanulate. The highest genetic distance was found between P. kurdica and *P. physocalyx*, with a genetic distance value of 0.625 within subsection Gymnophlomis in the present study (Table 2). While *P. kurdica* has either glandular or eglandular specimens, P. physocalyx has only eglandular specimens. The shape of the base of basal and cauline leaves is cordate in P. kurdica and cuneate in P. physocalyx. Number of flowers per verticillaster varies from 6 to 10 in P. kurdica and from 4 to 6 in *P. physocalyx*.

The third group included *P. tuberosa* from section *Phlomoides* and *P. samia* from section *Phlomis* subsection *Oxyphlomis*. *P. tuberosa* and *P. samia* have pink or purple corolla color in common. The fourth group contained taxa from section *Phlomis* subsection *Oxyphlomis*: *P. pungens* var. *pungens*, *P. pungens* var. *laxiflora*, *P. pungens* var. *hirta*, *P. pungens* var. *laxiflora*, *P. pungens* var. *hirta*, *P. pungens* var. *hispida*, *P. pungens* var. *seticalycina*, *P. rigida* and *P. integrifolia*. The shape of the base of *P. tuberosa* and *P. samia* is cordate-sagittate, but cuneate, truncate or obtuse in the species of the fourth group. Both

P. tuberosa and P. samia have a branched single stem; however, species in the fourth group have many stem above the soil except for P. rigida, which has a single stem in generally, but a branched stem occasionally. Also, P. tuberose and P. samia have bigger cauline leaves than the species in the fourth group. In species of section Phlomis subsection Oxyphlomis bracteoles are rigid and subulate in shape; 9-40 in number and 10-26 mm in length (Huber-Morath, 1982). Number of verticillasters in these species varies 2 to 6 in general (Huber-Morath, 1982). Within the fourth group, no protein variability was observed between P. pungens var. pungens and P. pungens var. laxiflora, and between P. pungens var. hispida and P. pungens var. seticalycina. Calyx teeth length is the only morphological diagnostic which discriminates P. pungens var. pungens from P. pungens var. laxiflora. Calyx teeth are 1/2 as long as calyx tube or longer in var. pungens, but they are 1/3-1/5 as long as calyx tube in var. laxiflora (Huber-Morath, 1982). Morphological characters which differentiate P. pungens var. hispida from P. pungens var. seticalycina are adaxial suface of leaves and hairy structure on the stem. Adaxial surface of leaves is subglabrose in var. seticalycina but stellate-tomentose in var. hispida, and stem is covered with sparsely undivided hairs in var. seticalycina; however, in var. hispida it is covered with dense indumentum of undivided hairs, especially near nodes (Huber-Morath, 1982). P. samia was found to be the most distant species to both P. pungens var. pungens and P. pungens var. laxiflora, with a genetic distance value of 0.769 within the subsection Oxyphlomis in the present analysis (Table 2). While P. samia has glandular hairs on stem, leaves and the floral regions such as bracteoles and calyx, P. pungens is eglandular. The shape of the base of the cauline leaves is cordate in *P. samia* and cuneate in *P. pungens*. Flowers are shortly pedicellated in *P. samia* and sessile in *P. pungens*. Number of flowers per verticillaster varies from 10 to 18 in P. samia and from 6 to 10 in P. pungens. Calyx length varies from 8 to 18 mm in P. pungens and from 18 to 23 mm in P. samia.

In the present study, P. tuberosa, the only member of section *Phlomoides* in the Flora of Turkey, was expected to out group with the species of section Phlomis, but it clustered with subsection Oxyphlomis. This position of P. tuberosa could be understandable when considering that this species and species of subsection Oxyphlomis have pink or purple corolla color in common. Nevertheless, the inefficiency of section discrimination in the genus might be due to insufficient number of species sampled from section Phlomoides and thus, the number of species sampled from section Phlomoides need to be increased to examine whether the sectional division of the genus is supported by the SDS-PAGE analysis of seed proteins in future studies. On the other hand, the grouping pattern of the taxa in the present study supports the view of Huber-Morath (1982) in which section Phlomis was delimited in three subsections as Dendrophlomis, Gymnophlomis and Oxyphlomis. However, the taxa of subsection Gymnophlomis are grouped with the taxa of subsection Dendrophlomis in the

dendrogram. This topology indicates that subsection *Dendrophlomis* is more closely related to subsection *Gymnophlomis* than subsection *Oxyphlomis*. This result is also in agreement with the morphological data that both subsections have yellow flower color in comparison to subsection *Oxyphlomis* that have purple or pink flower color. Overall, the results of the present study indicate that seed proteins in the genus *Phlomis* are useful characters to differentiate between species and to construct phylogenies for the assessment of evolutionary species relationships.

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REFERENCES

ALBALADEJO, R.G.; APARICIO, A. and SILVESTRE, S. Variation patterns in the *Phlomis x composite* (*Lamiaceae*) hybrid complex in the Iberian Peninsula. *Botanical Journal of Linnean Society*, May 2004, vol. 145, no. 1, p. 97-108.

ALBALADEJO, Rafael G.; AGUILAR, Javier F.; APARICIO, Abelardo and FELINER, Gonzalo N. Contrasting nuclear-plastidial phylogenetic patterns in the recently diverged Iberian *Phlomis crinita* and *P. lychnitis* lineages (*Lamiaceae*). *Taxon*, November 2005, vol. 54, no. 4, p. 987-998.

ALBALADEJO, Rafael G. and APARICIO, Abelardo. Population genetic structure and hybridization patterns in the Mediterranean endemics *Phlomis lychnitis* and *P. crinita (Lamiaceae)*. *Annals of Botany*, October 2007, vol. 100, no. 4, p. 735-746.

ALVAREZ, J.B.; MORAL, A. and MARTIN, L.M. Polymorphism and genetic diversity for the seed storage proteins in Spanish cultivated einkorn wheat (*Triticum monococcum* L. ssp. *monococcum*). *Genetic Resources and Crop Evolution*, August 2006, vol. 53, no. 5, p. 1061-1067.

AZIZIAN, D. and MOORE, D.M. Morphological and palynological studies in *Phlomis L., Eromostachys Bunge* and *Paraphlomis Prain (Labiatae). Botanical Journal of Linnean Society*, June 1982, vol. 85, p. 225-248.

BENTHAM, G. Labiatarum Genera et species. London, 1834, p. 323-644.

BHARGAVA, A.; RANA, T.S.; SHUKLA, S. and OHRI, D. Seed protein electrophoresis of some cultivated and wild species of *Chenopodium*. *Biologia Plantarum*, December 2005, vol. 49, no. 4, p. 505-511.

BUSSELL, John D.; WAYCOTT, Michelle and CHAPPILL Jennifer A. Arbitrarily amplified DNA markers as characters for phylogenetic inference. *Perspectives in*

Yüzbaşıoğlu, E. et al.

Plant Ecology, Evolution and Systematics, March 2005, vol. 7, no. 1, p. 3-26.

CHERDOUH, A.; KHELIFI, D.; CARRILLO, J.M. and NIETO-TALADRIZ, M.T. The high and low molecular weight glutenin subunit polymorphism of Algerian durum wheat landraces and old cultivars. *Plant Breeding*, August 2005, vol. 124, no. 4, p. 338-342.

FELSENSTEIN, J. *PHYLIP* (*Phylogeny Inference Package*) version 3.6a3. Distributed by the author. Seattle, Washington; Department of Genome Sciences, University of Washington, 2002.

GHAFOOR, Abdul; AHMAD, Zahoor; QURESHI, Afsari S. and BASHIR, Muhammad. Genetic relationship in *Vigna mungo* (L.) Hepper and *V. radiata* (L.) R. Wilczek based on morphological traits and SDS-PAGE. *Euphytica*, February 2002, vol. 123, no. 3, p. 367-378.

HEDGE, I.C. Labiatae of South-West Asia: diversity, distribution and endemism. *Proceedings of the Royal Society of Edinburgh*, 1986, vol. 89B, p. 23-35.

HUBER-MORATH, A. *Phlomis* L. In: DAVIS, P.H. ed. *Flora of Turkey and East Aegean Island*. Edinburgh; Edinburgh University Press, 1982, vol. 7, p. 102-126.

IGREJAS, G.; GUEDES-PINTO, H.; CARNIDE, V. and BRANLARD, G. Seed storage protein diversity in triticale varieties commonly grown in Portugal. *Plant Breeding*, 1999, vol. 118, no. 4, p. 303-306.

JHA, S.S. and OHRI, D. Comparative study of seed protein profiles in the genus *Pisum*. *Biologia Plantarum*, December 2002, vol. 45, no. 4, p. 529-532.

KARIHALOO, J.M.; KAUR, M. and SINGH, S. Seed protein diversity in *Solanum melongena* L. and its wild and weedy relatives. *Genetic Resources and Crop Evolution*, December 2002, vol. 49, no. 6, p. 533-539.

KIRSTEN, Janet H.; DAWES, Clinton J. and COCHRANE, Bruce J. Randomly amplified polymorphism detection (RAPD) reveals high genetic diversity in *Thalassia testudinum* banks ex König (Turtlegrass). *Aquatic Botany*, August 1998, vol. 61, no. 4, p. 269-287.

LAEMMLI, U.K. Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature*, August 1970, vol. 227, no. 5259, p. 680-685.

MIRALI, N.; EL-KHOURI, S. and RIZQ, F. Genetic diversity and relationships in some *Vicia* species as determined by SDS-PAGE of seed proteins. *Biologia Plantarum*, December 2007, vol. 51, no. 4, p. 660-666.

MOENCH, Conrad. *Methodus plantas Horti botanici et agri Marburgensis, a straminum situ describendi*. Marburgi Cattorum, Marburg, Germany. 1794.

OLIVIERI, Erna; HERBERT, Ben and RİGHETTİ, Pier Giorgio. The effect of protease inhibitors on the two dimensional electrophoresis pattern of red blood cell membranes. *Electrophoresis*, February 2001, vol. 22, no. 3, p. 560-565

PANIGRAHI, J.; KUMAR, D.R.; MISHRA, M.; MISHRA, R.P. and JENA, P. Genomic relationships among 11 species in the genus *Cajanus* as revealed by seed protein (albumin and globulin) polymorphisms. *Plant Biotechnology Reports*, April 2007, vol. 1, no. 2, p. 109-116.

RAMAGLI, Louis S. and RODRIGUEZ, Lewis V. Quantitation of microgram amounts of protein in two dimensional polyacrylamide gel electrophoresis sample buffer. *Electrophoresis*, April 1985, vol. 6, no. 11, p. 559-563.

ROHLF, F.J. *NTSYS-pc numerical taxonomy and multivariate analysis system version 1.7.* New York; Exeter Publications, 1992.

ROUT, Anusuya and CHRUNGOO, Nikhil K. Genetic variation and species relationships in Himalayan buckwheats as revealed by SDS-PAGE of endosperm proteins extracted from single seeds and RAPD based DNA fingerprints. *Genetic Resources and Crop Evolution*, June 2007, vol. 54, no. 4, p. 767-777.

STOILOVA, T.; CHOLAKOVA, N. and MARKOVA, M. Variation in seed protein and isoenzyme patterns in *Cucurbita* cultivars. *Biologia Plantarum*, September 2006, vol. 50, no. 3, p. 450-452.

SYROS, T.; YUPSANIS, T. and ECONOMOU, A. Fractionation and electrophoretic patterns of storage proteins of *Ebenus cretica*. A preliminary survey as a tool in taxonomy. *Biologia Plantarum*, April 2003, vol. 46, no. 3, p. 435-443.

TAYLOR, J.M. *Phlomis, the neglected genus, a guide for gardeners and horticulturists.* National Council for the Conservation of Plants and Gardens, Westbury-on-severn; Gloucester, England, 1998, p. 3-87. ISBN 0953241300.

YUZBASIOGLU, E.; ACİK, L. and OZCAN, S. Seed protein diversity among lentil cultivars. *Biologia Plantarum*, March 2008a, vol. 52, no. 1, p. 126-128.

YUZBASIOGLU, Ertugrul; DADANDI, Mehmet Y. and OZCAN, Sebahattin. Natural hybridization between *Phlomis lycia* D. Don x *P. bourgaei* Boiss. (*Lamiaceae*) revealed by RAPD markers. *Genetica*, May 2008b, vol. 133, no. 1, p. 13-20.

YUZBASIOGLU, Ertugrul and DADANDI, Mehmet Y. Phylogenetic relationships among species of the subsection *Dendrophlomis* Bentham. *Electronic Journal of Biotechnology*, October 2008, vol.11, no. 4. VLADOVA, R.; PANDEVA, R. and PETCOLICHEVA, K. Seed storage proteins in *Capsicum annuum* cultivars. *Biologia Plantarum*, August 2000, vol. 43, no. 2, p. 291-295.