The Phytochemical constituents and the effects of methanol extracts of Phyllanthus amarus leaves (kidney stone plant) on the hormonal parameters of Male guinea pigs.

1OBIANIME A.W; 2UCHE, F.I.

1Department of Pharmacology University of Port Harcourt
2 Department of Pharmacognosy and Ethnotherapy University of Port Harcourt, Nigeria.
Email: uchefideliaijeoma@yahoo.com. 08037066891

ABSTRACT: The effects of the methanolic extracts of the leaves of Phyllanthus amarus on the hormonal parameters of male Guinea pigs were investigated. The phytochemical screening of the leaves of Phyllanthus amarus was also carried out. The hormonal parameters investigated are testosterone, Leutinizing and Follicle stimulating hormone. The methanolic extract of the Phyllanthus amarus leaves (50-800mg/kg) caused a statistically significance increase (P < 0.001, ANOVA) in the level of Testosterone of the male Guinea pigs, from 2.3 ± 0.06 to 3.9 ± 0.05, 4.3 ± 0.6 and 2.8 ± 0.6 after the 7, 14th and 21st day of the administration of the extracts respectively. The highest increase was obtained after 14th day of treatment (4.3 ± 0.05). These effects were very comparable to the effects of Vitamin E on the testosterone of male Guinea pigs, which were obtained to be 3.0 ± 0.01, 3.1 ± 0.16 and 2.4 ± 0.30 for 7, 14th and 21st day respectively. These effects were dose- and time- dependents. The optimum effects (4.3 ± 0.05) were obtained at 400mg/kg of Phyllanthus amarus.

Furthermore, the methanol extracts of Phyllanthus amarus (800mg/kg) caused an insignificant change in the levels of Leutinizing (LH) and Follicle stimulating (FSH) hormones from 3.1±0.22 and 1.6±0.50 to 3.0±0.08 and 1.5±0.13 respectively. These effects were also comparable to the effects of Vitamin E on these hormones.

Finally, the phytochemical screening of the leaves of Phyllanthus amarus revealed the presence of flavonoids, tannins, alkaloids, terpenoids, steroids, saponins and cardiac glycosides. This may support or justify the claims on the use of the aerial part of this plant by traditional medicine practitioners to increase/improve libido and reproductive function in men. Although further studies need to be done to investigate the contribution of the seeds of this plant in the improvement of libido in men; also to isolate and characterize the active principles in the leaf extracts.

MATERIALS AND METHODS
All the chemicals used were of analar grade.

Plant Material
The leaves of Phyllanthus amarus were collected from the local garden within the premises of University of Port Harcourt in June 2008. The plant was identified and authenticated by Edwin Nwosu of department of Botany herbarium, University of Port Harcourt. Voucher specimen was maintained at the Herbarium.

The fresh leaves collected were air- dried for 10days, until a constant weight was attained.

Preparation of Extract
The dried leaves of Phyllanthus amarus were pulverized (100g). The crude drug was extracted with methanol using Soxhlet extraction method. The solid residue obtained was kept in a capped container in a refrigerator. Different concentrations of the extract were reconstituted from this stock.

Experimental animals
The male guinea pigs were collected from the animal house of University of Port Harcourt. The weight of the animals ranges from 300-600g. The animals were

* Corresponding author: Obianime A.W
allowed to acclimatize with the new environment for seven days before the experiment. They were housed in a cage of five animals per cage and were adequately feed throughout the experiment.

**Phytochemical screening**

Chemical tests were carried out on the methanolic extracts and on the powdered specimens using standard procedures to identify the constituents (Trease and Evans, 1989; Harborne, 1973) by characteristic colour changes as described by Sofowara, (1993); Odebedy and Sofowara, (1978).

**Hormonal assay**

The animals were grouped into 10 groups of five animals per group. The animals from different groups were given diethyl ether anesthesia and dissected. Their respective blood samples were collected in lithium heparinized tubes.

**Leutnanizing hormone (LH) and Follicle stimulating hormone (FSH) assay**

In the assay of LH and FSH, 50ml of standard or test sample was measured into appropriate well. 100ml of enzyme conjugate reagent was added into the well. This was gently mixed for 10 seconds and incubated at room temperature for 45 minutes. The incubated mixture was removed by flicking the plate contents into the well and washed 5 times with water. 100ml of tetra methyl was added to the incubated mixture at room temperature and allowed to react for 20 minutes. The reaction was stopped by addition of 100ml of stop solution to the well and readings were taken at 450nm within 15 minutes.

Concentration of the test (A) was calculated as follows:

\[ A = \frac{\text{Absorbance of test}}{X} \times \frac{\text{Concentration of standard}}{\text{Absorbance of standard}} \]

**Testosterone Enzyme immuno assay**

This was carried out in three stages namely: Reaction of antibody with serum testosterone and testosterone label, Magnetic solid phase separation step and Colour development step.

In the reaction of antiserum with serum testosterone and testosterone label, 50ul of test blood sample was pip pipetted into different tubes. The testosterone blocking reagent, diluted testosterone label and testosterone antiserum (100ul) were added to the test tube, covered and vortex mixed.

**Magnetic separation reagent; reaction**

100ul of testosterone separation reagent was added to different test tubes, covered and vortex mixed. The tubes were incubated in water bath at 37°C for 30 minutes.

The assay tubes were removed from water bath and placed on a magnetic base. The rack of tubes was kept upright in magnetic separation for 5 minutes after which the supernatant liquid from all the tubes were decanted. Then the tubes were changed from upright position and remove from the magnetic base.

**Washing step**

50ul of dilute testosterone enzyme immune assay (EIA) wash buffer was added to different tubes and vortex mixed. The rack of tubes was placed on a magnetic base. The tubes were kept upright in the magnetic separation for 5 minutes. The supernatant liquid were decanted from all the tubes and the separator was returned to an upright position. The rack of tubes was removed from the magnetic base. The whole process was repeated. This process is essential to remove all unbound components.

**Colour development step**

500ul of substrate solution was added to different test tubes, covered and vortex mixed. The tubes were transferred to 37°C water bath and incubated for 6 minutes. The tubes were removed from the bath and 1ml of EIA stop buffer was added to the different tubes and mixed. The rack of tubes was placed onto a magnetic base and the tubes were kept upright in magnetic separation for 10 minutes. The absorbance of the test sample and standard were recorded spectrophotometrically and compared with the blank.

**Statistical analysis**

Data were expressed as mean ± standard error of mean (S.E.M). Results were subjected to statistical analysis using one way analysis of variance (ANOVA). P < 0.05 was accepted as significance.
RESULTS AND DISCUSSION

*** Represents P < 0.001 level of significance (ANOVA).

LHP represents Leutenizing hormone level on treatment with *P. amarus*;
LHC Leutenizing hormone level, control;LHE Leutenizing hormone level on treatment with Vitamin E;FSHP Follicle stimulating hormone on treatment with *P. amarus*; FSHC Follicle stimulating hormone, control;FSHE Follicle stimulating hormone on treatment with Vitamin E;TP Testosterone level on treatment with *P. amarus*; TC Testosterone level, control; TE Testosterone level on treatment with Vitamin E.

Table 1: The dose-dependent effects of methanolic extracts of the leaf of *Phyllanthus amarus* on the hormonal parameters of male guinea pigs.

<table>
<thead>
<tr>
<th>Treatment (mg/kg)</th>
<th>Leutenizing Hormone (mlu/L)</th>
<th>Follicle Stimulating Hormone (mlu/L)</th>
<th>Testosterone (ng/ml)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>3.1 ± 0.22</td>
<td>1.5 ± 1.5</td>
<td>2.3 ± 0.6</td>
</tr>
<tr>
<td>P.a. 50</td>
<td>3.1 ± 0.28</td>
<td>1.5 ± 0.17</td>
<td>*2.6 ± 0.42</td>
</tr>
<tr>
<td>P.a.100</td>
<td>3.1 ± 0.25</td>
<td>1.4 ± 0.01</td>
<td>*2.8 ± 0.30</td>
</tr>
<tr>
<td>P.a.200</td>
<td>3.1 ± 0.01</td>
<td>1.5 ± 0.04</td>
<td>* *3.4 ± 0.45</td>
</tr>
<tr>
<td>P.a.400</td>
<td>3.0 ± 0.15</td>
<td>1.5 ± 0.02</td>
<td>**4.3 ± 0.48</td>
</tr>
<tr>
<td>P.a.800</td>
<td>3.1 ± 0.08</td>
<td>1.5 ± 0.13</td>
<td>* *3.9 ± 0.56</td>
</tr>
<tr>
<td>Vit. E (500IU)</td>
<td>2.7 ± 0.01</td>
<td>1.4 ± 0.01</td>
<td>*3.0 ± 0.01</td>
</tr>
</tbody>
</table>

Values are expressed as mean ± SEM of five observations (n = 5).
* represents significant values at P < 0.05 and ** significant values at P < 0.001 (ANOVA). P.a means *Phyllanthus amarus*.

Table 2: Phytochemical Screening

<table>
<thead>
<tr>
<th>Phytochemicals</th>
<th>P.amarus</th>
</tr>
</thead>
<tbody>
<tr>
<td>Alkaloids</td>
<td>+</td>
</tr>
<tr>
<td>Flavonoids</td>
<td>+</td>
</tr>
<tr>
<td>Terpenoids</td>
<td>+</td>
</tr>
<tr>
<td>Saponins</td>
<td>+</td>
</tr>
<tr>
<td>Tannins</td>
<td>+</td>
</tr>
<tr>
<td>Steroid</td>
<td>+</td>
</tr>
<tr>
<td>Resins</td>
<td>-</td>
</tr>
<tr>
<td>Cardiac glycosides</td>
<td>+</td>
</tr>
</tbody>
</table>

+ = present
- = absent

* Corresponding author: Obianime A.W
The Phytochemical constituents and the effects of methanol extracts of Phyllanthus amarus leaves (kidney stone plant) on .........

Table 3: The comparative effects of the time – dependent effects of methanolic extracts of the leaf of Phyllanthus amarus and Vitamin E, overtime on the hormonal parameters of the male guinea pigs.

<table>
<thead>
<tr>
<th>Treatment Period (400mg/kg)</th>
<th>Leutenizing Hormone (mlu/L)</th>
<th>Follicle Stimulating Hormone (mlu/L)</th>
<th>Testosterone (ng/ml)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>3.1 ± 0.22</td>
<td>1.6 ± 1.50</td>
<td>2.3 ± 0.06</td>
</tr>
<tr>
<td>Vit. E (500IU)</td>
<td>2.7 ± 0.01</td>
<td>1.4 ± 0.01</td>
<td>* 3.0 ± 0.01</td>
</tr>
<tr>
<td>P.a at 7 days</td>
<td>3.0± 0.15</td>
<td>1.5 ± 0.02</td>
<td>* *3.4 ± 0.50</td>
</tr>
<tr>
<td>Vit. E</td>
<td>3.1 ± 0.16</td>
<td>1.5 ± 0.14</td>
<td>* 2.9± 0.16</td>
</tr>
<tr>
<td>P.a at 14 days</td>
<td>3.1 ± 0.11</td>
<td>1.4 ± 0.04</td>
<td>* *3.4± 0.60</td>
</tr>
<tr>
<td>Vit. E</td>
<td>3.1± 0.13</td>
<td>1.5 ± 0.05</td>
<td>2.1± 0.30</td>
</tr>
<tr>
<td>P.a at 21 days</td>
<td>3.2± 0.05</td>
<td>1.4± 0.01</td>
<td>2.4± 0.45</td>
</tr>
<tr>
<td>Vit. E</td>
<td>2.5±0.01</td>
<td>1.4± 0.02</td>
<td>2.6± 0.01</td>
</tr>
<tr>
<td>P.a at 28 days</td>
<td>3.0 ± 0.03</td>
<td>1.4 ± 0.03</td>
<td>2.4± 0.50</td>
</tr>
</tbody>
</table>

Values are expressed as mean ± SEM of five observations (n = 5).
* represents significant values at P< 0.05 and ** significant values at P < 0.001 (ANOVA). P.a means Phyllanthus amarus.

This study shows the effects of methanolic extract of the leaves of phyllanthus amarus on the hormonal parameters of male guinea pigs. The results show that Phyllanthus amarus leaf extract causes an increases in the level of testosterone but has little or no effect on the levels of Leutinizing hormone (LH) and follicle stimulating hormone (FSH) ,(table1 and 3; fig 1).The increase in the level of testosterone was found to be statistically significant at P < 0.001 (ANOVA). The increase in the testosterone may be responsible for the effect of the aerial part of this plant as a libido enhancer of fertility agent as claimed by traditional medicine practitioners. This is so because optimum level of testosterone is required for normal sex drive in adult male and an increase in the level of testosterone can lead to an increase in the spermatozoa (Vander et al., 2001) and hence an increase in male fertility (Joy and Kuttan, 1998). The phytochemicals found present in the leaf of P. amarus include: flavonoids, tannins, saponins, alkaloids, terpenoids, steroids and cardiac glycosides. Flavonoids present in this plant has been shown to possess many pharmacological properties such as: anti-oxidant activities, anti-inflammatory activities, anti- cancer activities and anti- microbial effects hence, flavonoids may have a contributory effect to its fertility properties and other pharmacological effects the plant possesses( Joy and Kuttan, 1998; Kassuya et al., 2003; Adeneye, 2006). Flavonoids as an anti-oxidant, has a rejuvenating effects on cells or tissues, it is anti-aging hence can contribute substantially on the fertility effect of this plant. Alkaloids and tannins may also contribute to the plant’s effects as antimalarial, anti- diarrhea and analgesic agents. This study therefore, supports the claims on the folkloric use of the aerial part of this plant to improve libido and reproductive function in men. However, further study needs to be to investigate the actual contribution of the seed of this plant as a fertility agent and also to isolate, identify and characterize the active principle present in the leaf of this plant.

REFERENCE


Kokwaro, J.O (1976). Medicinal Plants of East Africa Literature Bureau. 95


* Corresponding author: ‘Obianime A.W
The Phytochemical constituents and the effects of methanol extracts of Phyllanthus amarus leaves (kidney stone plant) on ... 


* Corresponding author: ‘Obianime A.W