372

PROS AND CONS OF IMMUNOCHEMICAL AND ENZYMATIC METHOD IN THE DIAGNOSIS OF WILSON'S DISEASE

J. V. GNANOU, V. G. THYKADAVIL*, V. THUPPIL*

ABSTRACT

BACKGROUND: Immunochemical method of measuring Ceruloplasmin (Cp) levels for the diagnosis of Wilson's disease has replaced enzymatic method for the main reason of being more sensitive and quantitative. **SETTINGS AND DESIGN:** In this study, we compared both the methods for various factors such as sensitivity, specificity and the time consumed in the diagnosis of Wilson's disease. **MATERIALS AND METHODS:** Serum samples from patients (n=33) with a provisional diagnosis of Wilson's disease were analyzed for Cp levels by enzymatic copper oxidase method and immunochemical method using polyclonal antibodies specific to Cp embedded in agar. **STATISTICAL ANALYSIS:** Pearson's regression analysis was performed to compare the two methods. **RESULTS:** The mean Cp obtained by immunochemical method is 5.87 mm \pm 1.17 and by enzymatic method, it is 0.37 (Optical Density) \pm 0.136. Pearson's Regression analysis of the measurements showed a good correlation with an 'r' value of 0.765 between the two methods. **CONCLUSION:** A good correlation indicated that these two tests are comparable and thus both these methods can be used together for a definitive and better diagnosis of Wilson's disease.

Key words: Ceruloplasmin, copper oxidase method, immunochemical method, Wilson's disease

Cp is a transport protein for copper and is secreted as apoCp. This protein takes up six to eight atoms of copper through a copper transport mechanism using ATP, producing holoCp.^[1] The disruption of the coppertransporting mechanism due to mutations (ATP7B) leads to a reduced excretion of copper and accumulation of copper in

Correspondence

Justin V. Gnanou, C/o Dr. Vinod George, Dept. of Biochemistry, St. John's Medical College, Koranamgala, Bangalore - 560 034, India. E-mail: gnanou_j@yahoo.com hepatolenticular cells causing Wilson's disease. More than 200 known mutations have been characterized and documented.^[2] High level of suspicion in pediatric populations presenting with neuro-hepatic problems prompts clinicians to monitor their levels of Cp. The diagnosis of Wilson's disease is made by measuring the Cp levels, which will be lower, along with clinical signs and elevated liver enzymes.^[3] There are two methods for estimating Cp in blood: copper oxidase method and immunochemical method. Copper oxidase method is an enzymatic method and estimates the holoCp, while the immunochemical method estimates

the combined apoCp and holoCp. Hence it is more appropriate to estimate the holoCp levels rather than apoCp and holoCp together in the diagnosis of Wilson's disease. Also, since Cp is an acute phase reactant protein. inflammation due to any cause can increase the Cp levels. Thus immunochemical method has lower specificity than copper oxidase method for the diagnosis of Wilson's disease.^[4] Hence using immunochemical method in patients with Wilson's disease presenting with inflammation or infection, normal or high Cp levels can be obtained.[5,6] This problem is overcome by using copper oxidase method, which measures only the holoCp levels. However, copper oxidase method is a qualitative method and has low sensitivity.^[7] The aim of this study was to study the pros and cons of these two methods, immunochemical method and copper oxidase method, in patients suspected with Wilson's disease.

MATERIALS AND METHODS

Subjects

This study was a prospective study and was conducted over a period of 6 months-from January to June 2005. The study population was selected using an inclusion criterion of pediatric patients between 1 year and 15 years of age, with nonspecific liver diseases in whom Wilson's disease was the differential diagnosis. Patients with specific liver diseases such as acute or chronic hepatitis and adult patients were excluded from the study. Since this study was restricted to a period of 6 months, all patients who fitted the inclusion criteria were selected for the study. Thus the size of the study population was 33. Of these 33 patients, 31 were male and 2 were female. Two milliliters of blood was collected by standard venipuncture procedure and samples were immediately centrifuged at 5,000 rpm at 25°C and serum was separated. The samples were collected on different days for a period of 6 months as and when patients were recruited. The serum samples were aliquoted into three vials. The first two vials were coded and they were immediately analyzed for Cp levels using both the immunochemical method and the copper oxidase method. The third serum vial was immediately stored at - 20°C for future and repeat analysis. The lab staff members were blinded for this analysis.

Ethical clearance

Ethical approval was obtained from the Institutional Ethical Review Board of St. John's Medical College, Bangalore and all subjects gave informed consent.

Sample analysis

Immunochemical method was done using polyclonal antibodies specific to Cp embedded in agar (Biocientifica, SA Buenos Aires, Argentina). In this method, 10 µL of serum sample was added into the sample well in the agar. The sample was incubated in the agar medium at room temperature for about 12 h. After the incubation period, the Cp concentrations were calculated by measuring the diameter of the radial immunodiffusion by a calibrated scale.^[8] This measurement was then compared to a control run along with the sample and result given in mg/dl. The enzymatic oxidase activity of Cp was assayed using paraphenylenediamine in sodium acetate as substrate and the purple product was

University of Seychelles American Institute of Medicine, MAHE, Seychelles, *Department of Biochemistry and Biophysics, St. John's National Academy of Health Sciences, Johnnagar, Bangalore, India

373

374

measured spectrophotometrically at 530 nm.^[6,8] In this method, 100 μ L of serum sample was taken in a test tube and incubated with para-phenylenediamine in sodium acetate buffer for 15 min. After the 15-min incubation period at room temperature, the color produced was measured as optical density at 530 nm using a spectrophotometer. This value was then compared to a control/normal sample run along with the test with same lab conditions. If the optical density of the test was lower than the normal, then the test was interpreted as Cp deficiency. If the optical density was same or higher than the control/normal sample, the test was interpreted as normal Cp levels. The within-run CV% and betweenrun CV% of immunochemical method was 1.57 and 2.63% respectively and the withinrun CV% and between-run CV% for copper oxidase enzymatic method was 3.23 and 7.01% respectively. The lower limit of detection of immunochemical is 7 mg/dl.

Statistical analysis

Analysis was performed with SPSS (SPSS 10.1 for windows, SPSS Inc 2000). The two methods were compared using Pearson's regression analysis. For regression analysis, 'r' value greater than 0.5 was considered as good correlation.

RESULTS

The mean Cp obtained by immunochemical method was 5.87 mm \pm 1.17 and by

enzymatic method, it was 0.37 (optical density) \pm 0.136. Pearson's regression analysis of the measurements showed a good correlation with an 'r' value of 0.765 between the two methods [Table 1].

From the total observations, the predicted and residual values were plotted onto the X and Y axis to obtain a line-fit plot. Using this plot, a predictive equation was arrived at. For this study, the predictive equation is as follows: Optical density by enzymatic method = $0.0893 \times \text{millimeters}$ measured by immunochemical method - 0.1475 [Figure 1]. The 95% confidence limits of the regression analysis is given in Table 2.

DISCUSSION

In this study, we found a good correlation



Figure 1: Regression analysis of immunochemical method and enzymatic method for estimation of Cp levels. Comparative analysis of immunochemical method and enzymatic method for Cp. Immunochemical method (millimeters) is plotted on X-axis and enzymatic method (optical density) is plotted on Y-axis.

Table 1: Comparison of Cp levels by immunochemical and copper oxidase enzymatic methods

| Subjects | Cp level (mm) by immunochemical method | Cp level (optical density) by enzymatic oxidase method | r-value |
|-------------------|--|--|---------|
| Patients (n = 33) | 5.87 mm ± 1.17 | 0.37 (optical density) \pm 0.136 | 0.765 |

Table 2: Gives the 95% confidence limits of the regression analysis:

| | Upper 95% | Lower 95% |
|---------------|-----------|-----------|
| Intercept: | 0.01704 | -0.3121 |
| X variable 1: | 0.1167 | 0.0617 |

between the two methods - immunochemical method and enzymatic method - for the diagnosis of Wilson's disease.

The gold standard method for the diagnosis of Wilson's disease is through mutation analysis. The only drawback of this method is the cost-effectiveness.^[2] Mutation analysis costs approximately US \$ 200-250, while the immunochemical method costs approximately US \$ 10 and enzymatic method costs approximately US \$ 2. Immunochemical method is a sensitive and quantitative method but measures the total Cp (holo + apo Cp). Thus in patients with Wilson's disease presenting with infection,^[9] this test will give a false negative result. On the other hand, copper oxidase method is specific for the holoCp and does not measure the nonbound Cp. But this test has a disadvantage of being less sensitive and being a qualitative test. This test also is labor intensive and time consuming. Thus most labs prefer to use a faster, quantitative and easier method, which is the immunochemical method, for the estimation of Cp. We also found in our hospital through personal communications that clinicians were more inclined to trust a quantitative result than a qualitative one. Hence when using Cp assays for diagnosing Wilson's disease, the clinicians should be aware of the nonspecificity of the immunochemical method. Moreover, the lab should also take responsibility of informing the clinician about the method used and the

advantages and disadvantages of the method.

With due consideration of the above points, we analyzed Cp levels in patients with differential diagnosis for Wilson's disease by these two methods to see if they were comparable. We found that these two methods are comparable. Since they are comparable, both the methods can be used simultaneously. Thus by combining both these methods, the problem of sensitivity and specificity can be avoided.

The main drawback of this study was that the patients were not categorized based on inflammatory status. Also a definitive diagnosis of Wilson's disease using mutation analysis would have given better information of the usefulness of these assays in the diagnosis of Wilson's disease. The other main drawback of this study was that this study was restricted to pediatric population. In future, perhaps, it may be possible to include all the age groups in order to eliminate the role of age and sex in these assays.

To conclude, we found that enzymatic assay is the test of choice for the diagnosis of Wilson's disease, but combination of both immunochemical and enzymatic method would eliminate the disadvantages of both the methods.

ACKNOWLEDGMENT

All authors appreciate the efforts put in by Ms. Geetha S. for providing technical help in this study.

REFERENCES

1. Tanzi RE, Petrukhin K, Chernov I, Pellequer JL,

Wasco W, Ross B, *et al.* The Wilson disease gene is a copper transporting ATPase with homology to the Menkes disease gene. Nat Genet 1993;5:344-50.

- Petrukhin K, Fischer SG, Pirastu M, Tanzi RE, Chernov I, Devoto M, *et al.* Mapping, cloning and genetic characterization of the region containing the Wilson disease gene. Nat Genet 1993;5:338-43.
- Walshe JM. Diagnostic significance of reduced caeruloplasmin concentration in neurological disease. Mov Disord 2005;20:1658-61.
- Schilsky ML, Shneider B. Population screening for Wilson's disease. J Pediatr 2002;140:499-501.
- 5. Langner C, Denk H. Wilson disease. Virchows Arch 2004;445:111-8.
- 6. Steindl P, Ferenci P, Dienes HP, Grimm G, Pabinger I, Madl C, *et al.* Wilson's disease in patients presenting

with liver disease: A diagnostic challenge. Gastroenterology 1997;113:212-8.

- Macintyre G, Gutfreund KS, Martin WR, Camicioli R, Cox DW. Value of an enzymatic assay for the determination of serum ceruloplasmin. J Lab Clin Med 2004;144:294-301.
- Walshe JM. Clinical Investigations Standing Committee of the Association of Clinical Biochemists. Wilson's disease: The importance of measuring serum caeruloplasmin non-immunologically. Ann Clin Biochem 2003;40:115-21.
- Shifrine M, Fisher GL. Ceruloplasmin levels in sera from human patients with osteosarcoma. Cancer 1976;38:244-8.

Source of Support: Nil, Conflict of Interest: None declared.

Author Help: Reference checking facility

The manuscript system (www.journalonweb.com) allows the authors to check and verify the accuracy and style of references. The tool checks the references with PubMed as per a predefined style. Authors are encouraged to use this facility before submitting articles to the journal.

- The style as well as bibliographic elements should be 100% accurate to get the references verified from the system. A single spelling error or addition of issue number / month of publication will lead to error to verifying the reference.
- Example of a correct style Sheahan P, O'leary G, Lee G, Fitzgibbon J. Cystic cervical metastases: Incidence and diagnosis using fine needle aspiration biopsy. Otolaryngol Head Neck Surg 2002;127:294-8.
- Only the references from journals indexed in PubMed would be checked.
- Enter each reference in new line, without a serial number.
- Add up to a maximum 15 reference at time.
- If the reference is correct for its bibliographic elements and punctuations, it will be shown as CORRECT and a link to the correct article in PubMed will be given.
- If any of the bibliographic elements are missing, incorrect or extra (such as issue number), it will be shown as INCORRECT and link to possible articles in PubMed will be given.