Brain edema after intracerebral hemorrhage in rats: The role of inflammation

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**This article was supported by Hebei provicine, No.: 05276101D-27.

Background: Intracerebral hemorrhage (ICH) results in secondary brain edema and injury that may lead to death and disability. ICH also causes inflammation. It is unclear whether inflammation contributes to brain edema and neuron injury or functions in repairing the brain tissue. Aims: To understand the effect of inflammation in ICH, we have carried out an investigation on the various aspects and the dynamic changes of inflammation. Settings and Design: An ICH model was generated by injecting 50 µl autologous tail artery blood stereotactically into the right caudate nucleus of 30 rats, which were randomly divided into five ICH groups. Similarly, five Sham control groups were generated by inserting the needle to the right caudate nucleus of rats. Materials and Methods: Rat behavior was evaluated over the time course (6 h, 24 h, 48 h, 72 h and 7 d) in each group. The rats were then killed by administering an overdose of pentobarbital. Following the euthanasia, the brain water content, neuronal loss, glia proliferation, inflammatory infiltration and brain morphology of the rats were measured. Additionally, the expression of TNF-α, IL-6, ICAM-1, VEGF, NF-κB, C3 and CR2 was analyzed by immunohistochemistry. Statistical Analysis: The data were analyzed by student’s t test. Results: Rat brain water content increased progressively over the time course and reached its peak at 48h followed ICH. The maximum of inflammatory infiltrate (especially neutrophils) and immunopositive cells of TNF-α, IL-6 and NF-κB, were at 48h. The expression of C3 and CR2 reached their peaks at 48-72h, while the expression ICAM-1 and VEGF were at maximum at 72h followed ICH. Conclusions: The results suggested that the inflammatory cytokines, complement system and VEGF may have a function in the development of the brain edema and neuron injury followed ICH.

Key words: Animal models, brain edema, inflammation, intracerebral hemorrhage

Materials and Methods

ICH rat model

The Institutional Animal Care and use committee and the local experimental ethics committee have approved all experimental procedures. Sixty male Sprague-Dawley rats (13-15 weeks) were used. The rats were anesthetized with pentobarbital (40 mg/kg, IP) and positioned in a stereotaxic frame (SR-6N, Japan). A cranial burr hole (1 mm) was drilled on the right coronal suture 3.5 mm lateral to the midline. ICH was initiated in 30 rats stereotactically by infusing 50 µl autologous tail artery blood into the right caudate nucleus at 10 µl/min through a 26-gauge needle (coordinates: 0.2 mm anterior; 5.5 mm ventral and 3.5 mm lateral to bregma). Thirty rats served as sham controls with only a needle insertion. At the end the needle was removed, the burr hole was sealed with bone wax, the wound was sutured and the animal was
placed in a warm box with free access to food and water. Physiological parameters were maintained in the normal range in this experiment.

**Behavior tests**

Four motor behavior tests were carried out on rats by observers blinded to the experimental group or Sham group at time points 6h, 24h, 48h, 72h and seven days postoperation. (1) Longa behavioral test,\(^{[3]}\) measured the spontaneous contralateral circling and tumbling and was graded from 0 (no circling) to 4 (unconsciousness); (2) Berderson behavioral test,\(^{[4]}\) measured the palsy of the contralateral limbs and was graded from 0 (no palsy) to 3 (circling of the contralateral); (3) Beam walking test,\(^{[5]}\) measured the ability to walk on an 80-cm-long, 2.5-cm-wide wood beam and was graded from 0 for a rat that readily traversed the beam to 5 for a rat that was unable to move or fell off the beam; (4) Footfault asymmetry test,\(^{[6]}\) measured the ability to walk in a net (the areole was 2.3 x 2.3 cm).

**Brain water content measurement**

At time points 6h, 24h, 48h, 72h and seven days, after the neurobehavioral tests were done, each rat was killed by a pentobarbital overdose. The brain was quickly removed and placed on a cooled surface. The frontal pole (approximately 3 mm thick),\(^{[6]}\) the cerebellum and brainstem were removed. The cerebrum was coronally divided into three pieces by sectioning through the needle entry site and the midpoint of the posterior remnant, respectively. The first piece (2 mm thick) was cut ipsilateral and contralateral of ICH, the two sections were used for brain edema measurement. Each section was wrapped in preweighed aluminum foil and weighed to obtain the wet weight (WW), then dried for 72h in an oven at 110°C and weighed again to obtain the dry weight (DW). Brain water content was calculated as the percentage change using the following formula: \(\text{WW-DW}/\text{WW} \times 100\%\).

**Histological examination**

The other two coronal specimens from the cerebrum were used for histological examination. Briefly, they were perfused with 4% paraformaldehyde in 0.1 mol/L PBS (pH 7.4) for three days and then dehydrated and embedded in paraffin. Sections (5 µm) were stained with hematoxylin and eosin or stained by routine immunohistochemical methods. Primary antibodies used were TNF-α (diluted 1:100, Santa Cruz, USA), IL-6 (diluted 1:50, Boshide company, China), ICAM-1 (diluted 1:100, Zhongshan biology technology company, China), VEGF (diluted 1:150, Zhongshan biology technology company, China), NF-KB (diluted 1:200, Santa Cruz, USA), C3 (diluted 1:500, Sigma, USA) and CR2 (complement receptor type 2, CD21) (diluted 1:500, Santa Cruz, USA). The secondary antibodies, secondary biotinylated conjugates and dianinobenzidine were from the Vector ABC kit (Zhongshan biology technology company, China). The immunohistochemical labeling and detection were performed following the protocols recommended by the manufacturers.

**Statistical analysis**

All the values were shown as mean and standard deviation (means ± SD), statistical analysis software SPSS 11.0 was used for data analysis. ANOVA and t-tests were carried out for water content data and student’s t-test was carried out for the rest of the data. Differences were considered significant at \(P<0.05\).

**Results**

**High neurological deficit scores at 24h, 48h and 72h after operation**

The behavior tests showed that the maximum neurological deficit scores in both ICH and Sham groups were at 6h postoperation, a phenomenon caused by the anesthesia and injuries during the operation [Figures 1.1-1.4]. The neurological deficit score of the ICH group maintained a high level at 24h, 48h and 72h, but decreased to the lowest level at seven days when the rats had recovered from the operation. In contrast, the scores of Sham control rats were lower than that of the ICH rats at 24h, 48h and 72h. The differences of the four behavior tests between the two groups were statistically significant, the \(P\) values at 24h, 48h, 72h were 0.011, 0.008, 0.002 respectively for the Longa test [Figure 1.1], 0.003, 0.012 and 0.001 for the Berderson test [Figure 1.2], 0.02, 0.02 and 0.002 for the Beam Walking test [Figure 1.3], and 0.041, 0.021, 0.004 for the Footfault asymmetry test [Figure 1.4].

**Significant difference of brain water content between ICH and Sham rats at 48h, 72h**

Brain water content of the ICH and Sham groups showed no significant difference at 6h \(P=0.157\) and 24h \(P=0.830\) [Figure 2]. At 48h and 72h, the brain water content of both ICH ipsilateral and contralateral samples was significantly higher than those of Sham group \(P<0.001\), with maximum water content observed at 48h. Significant differences were also observed between the ipsilateral and contralateral samples at 48h \(P=0.0430\) and 72h \(P=0.049\).

**Significant changes in the brain tissue of rats in ICH group**

A spherical hematoma was observed in the caudate nucleus area at all time points after ICH. At time point 6h, a few rounded scattered neutrophils were found around the periphery of the hematoma. No morphological changes of neurons were observed. At 24h, brain edema around the hematoma was visible, the infiltrated inflammatory cells were mainly mononuclear and several neutrophils could be seen. At 48h, the brain edema around the hematoma was pronounced and the brain tissue was diffusent and necrotic [Figure 3]. The hematoma was surrounded by a compact band of cells including viable neutrophils, some cell debris, a few macrophages and rare clusters of intact erythrocytes. Degenerated neurons with vague nuclei and disappearing Nissl bodies and neuronophagia were also observed. At 72h, hematoma...
was surrounded by scattered glia cell containing hemosiderin, hyperplasia of glia and neovascularization. At time point seven days, the hematoma was resolving. The glia cell hyperplasia and neovascularization were abundant. Neutrophils were not present. In contrast, no hematoma or significant changes were observed in the brain tissue of rats from Sham group.

Expression of inflammatory factors

Positively stained neurons with a nucleus and normal morphology were counted as activated cells in three nonoverlapping regions under the microscope for the ipsilateral perihemotoma brain tissues. The expression of TNF-α [Figure 4], ICAM-1 [Figure 5], IL-6 [Figure 6] and NF-κB [Figure 7] after ICH were mainly in the cytoplasm of neurons and glia and reached their maximum at 48h or 72h. The expression of complement 3 [Figure 8] and CR2 [Figure 9] after ICH was up-regulated on the neurons, glia and endothelial cells in the perihemotoma. The expression of VEGF was up-regulated and maintained at high level after 48h, immunopositive cells were mainly neutrophils, neurons and endothelial cells [Figure 10].

Discussion

Animal model

Several experimental models of ICH have been described.\[8-10\]
Figure 4: Dynamic expression tendency of TNF-α in the brain tissues around hematoma in rats from the ICH and Sham groups at different time points

Figure 7: Dynamic expression tendency of NF-κB in the brain tissues around hematoma in rats from the ICH and Sham groups at different time points

Figure 5: Dynamic expression tendency of ICAM-1 in the brain tissues around hematoma in rats from the ICH and Sham groups at different time points

Figure 8: Dynamic expression tendency of complement 3 in the brain tissues around hematoma in rats from the ICH and Sham groups at different time points

Figure 6: Dynamic expression tendency of IL-6 in the brain tissues around hematoma in rats from the ICH and Sham groups at different time points

Figure 9: Dynamic expression tendency of CR2 in the brain tissues around hematoma in rats from the ICH and Sham groups at different time points

With modification of previous ones, here we developed an ICH rat model by sampling the blood from the tail artery instead of the femoral artery. The modifications made the model more reliable since the procedure had no effect on the neurobehavioral evaluation.

Brain water content and neurobehavioral evaluation

We demonstrated that the brain edema reached its peak between 48h and 72h and resolved at seven days after ICH, consistent with that of the neurobehavioral deficit scores. This suggested that there was a possible correlation between brain edema and motor behavior changes. Since four neurobehavioral tests were used for evaluation of the motor behavior, the neurobehavioral deficit scores should be more reliable.

Inflammatory cytokines: TNF-α, IL-6, ICAM-1 and NF-κB

Neutrophilic inflammation is in the vicinity of cerebral hematoma. Neutrophils release a variety of cytokines which might
play an important role in brain edema formation and aggravation, such as TNF-α, ICAM-1, IL-6 and NF-κB. It is also characterized by leukocyte behavior changes in the microvessels. Many leukocytes roll and adhere to the postcapillary venule and capillary walls and then these neutrophils infiltrate and migrate outside the vascular walls and move into the parenchyma. This inflammatory response is associated with expression of inflammatory mediators, including inflammatory cytokines, chemokines and adhesion molecules. In turn, infiltrated neutrophils release proteases and oxidases and result in secondary brain injury. Our results indicated that neutrophils around the hematoma were the most abundant at 48h to 72h postoperation; the amount of inflammatory cytokines such as IL-6 and TNF-α increased and reached their peaks at 48h after ICH. The expression of ICAM-1 was up-regulated, correlated with the action of IL-6 and TNF-α after ICH. Additionally, the immunopositive ICAM-1 neurons were the most abundant at 72h. ICAM-1 neurons might contribute to the adhesion of activated microglia to neurons, leading to the neuron injury and brain edema after ICH. Transcription factor NF-κB plays a key role in secondary impairments of the tissues around the hematoma by inducing the expression of various genes related to cell injury and apoptosis. Here we demonstrated that the expression of NF-κB was consistent with the time course of brain edema after ICH, suggesting that NF-κB may worsen the brain edema and brain injury.

**Complement system: C3 and CR2**

C3 is an indicator of complement activation. C3d is a degradation product of activated C3. CR2 is a receptor for C3d. It belongs to a family of complement regulatory proteins and is known for its bridging function at the intersection of innate host defense and acquired humoral immunity. Possibly by the adjuvant effect of C3d, the C system (C3 and CR2) may be involved in selecting antigens for recognition by the acquired immune system. This selecting antigen function is an immunity-augmenting function. The enhancement magnitude of antigen attachment of C3d might be 10,000-fold, which is far greater than that of complete Freund’s adjuvant. It showed that local activation of complement in the brain is of pathophysiological significance in both degenerative and inflammatory neurological diseases including cerebrovascular disorders. Complement depletion significantly reduced edema formation at both 24 and 72h after ICH. We demonstrated the up-regulation of C3 and CR2 protein levels in response to ICH in rat brain, suggesting C3 and CR2 may play a role in the development of inflammation, brain edema and brain injury after ICH.

**VEGF**

The results showed that the VEGF in rat brain after ICH was up-regulated. The up-regulation of VEGF correlates with blood-brain barrier breakdown and vasogenic brain edema formation. VEGF expression reached its peak at 72h. However, many positive cells, small vessel and vessel-like structures were still observed at seven days, suggesting that VEGF promoted the late stage of vasogenic edema. VEGF accelerates the breakdown of endothelial cells and the basilar membrane, which could subsequently increase the leukocyte infiltration and aggravate brain edema. VEGF might stimulate the angiogenic response and neovascularization.

**Conclusion**

This study characterized the brain edema, neurobehavioral deficit and inflammatory response after ICH in rat. Our results showed that cytokines, complement system and blood-brain barrier breakdown might play a role in the development of the brain edema and neuron injury after ICH. Our work will help to shed light on understanding of how the brain modulates inflammatory injury responses.

**Acknowledgment**

We would like to thank technician Hongran Wu and Biaofen Jin for their technical assistance.

**References**

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Accepted on 07-08-2006

Source of Support: Nil. Conflict of Interest: None declared.