Chronic Trypanosoma cruzi-elicted cardiomyopathy: from the discovery to the proposal of rational therapeutic interventions targeting cell adhesion molecules and chemokine receptors - how to make a dream come true

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One hundred years ago, Carlos Chagas discovered a new disease, the American trypanosomiasis. Chagas and co-workers later characterised the disease’s common manifestation, chronic cardiomyopathy, and suggested that parasitic persistence coupled with inflammation was the key underlying pathogenic mechanism. Better comprehension of the molecular mechanisms leading to clinical heart afflictions is a prerequisite to developing new therapies that ameliorate inflammation and improve heart function without hampering parasite control. Here, we review recent data showing that distinct cell adhesion molecules, chemokines and chemokine receptors participate in anti-parasite immunity and/or detrimental leukocyte trafficking to the heart. Moreover, we offer evidence that CC-chemokine receptors may be attractive therapeutic targets aiming to regain homeostatic balance in parasite/host interaction thereby improving prognosis, supporting that it is becoming a non-phantasious proposal.

Key words: Chagas disease - Trypanosoma cruzi - cardiomyopathy - chemokine - chemokine receptors - cell adhesion molecules

In 1909, Carlos Chagas realised a unique medical finding describing a new disease known as American trypanosomiasis, as well as its vector and causative agent, the haemoflagellate protozoan Trypanosoma cruzi (Chagas 1909). This discovery initiated a movement to characterise the disease’s clinical manifestations and epidemiological significance (Laranja et al. 1956). One hundred years after its discovery, Chagas disease is still epidemiologically relevant, afflicting 12-14 million people in 18 endemic countries in Latin America (Cora 2007, Dias 2007). Several hundred thousand T. cruzi-infected individuals may also be living in North America, Europe and other countries as a result of immigration to these areas in search of a better life (Schmunis 2007), demonstrating that Chagas disease is a far-reaching public health issue.

Chagas’ (1909, 1911) initial studies described the peculiar clinical symptoms associated with American trypanosomiasis, including cardiac excitability disturbance, extrasystoles, abnormal conduction and complete heart blockage. In 1916, Chagas compiled the histopathological findings from acute and chronic cases of human as well as experimental T. cruzi infection. Based on these data, he proposed that the pathogenic mechanism underlying cardiac symptoms was intense parasitism coupled with inflammation during the acute phase and parasite persistence, “although in small numbers inside cardiomyocytes making them difficult to find”, in parallel with inflammation during chronic infection (Fig. 1). Curiously, Chagas described the presence of amastigote forms of the parasite in the heart tissue of chronically infected individuals (Fig. 1). Assuming that inflammation was not proportional to the small numbers of parasite nests, he claimed that the inflammatory lesions resembled “an allergic process”. However, Chagas noted that in “most of the chronic cases with predominant cardiac signs a large number of protozoa is observed in the myocardium and the inflammatory reactions of the organ is also more intense” (Chagas 1916).

During the following years, Chagas and Villela (1922) adopting a new non-invasive procedure for recording the heart’s electrical activity (electrocardiography, or ECG) and recently described parameters to characterise heart abnormalities defined the precise cardiologic profiles of acutely and chronically T. cruzi-infected patients. However, three decades passed before for Chagas’ original observations of cardiac disturbances during chronic T. cruzi infection were reproduced in experimentally infected dogs (Laranja et al. 1950, 1956) and confirmed in a large number of patients (Laranja et al. 1956). Chagas’ detection of parasite nesting in cardiomyocyte or phagocytized by macrophages in all studied chronic chagasic
cardiomyopathy (CCC) patients (Chagas 1916, Chagas & Vilella 1922), although “a time-consuming activity” was also confirmed by Laranja et al. (1956). Moreover, cardiac histopathological abnormalities were characterised as consisting of diffuse and focal heart inflammatory foci chiefly composed of lymphocytes, plasma cells and macrophages, as well as eosinophils and neutrophils. Focal and diffuse fibrosis of the myocardium was also evidenced in all cases, its intensity varying from case to case and from one area to another within the same case (Laranja et al. 1956). Together, these findings provide a firm basis for defining CCC as a distinct clinical and anatomical entity.

Intriguingly, T. cruzi parasites were not observed in the cardiac tissue lesions of most patients with chronic Chagas disease for decades. Scientists began to ascribe a major role to autoimmune reactions in CCC pathogenesis (Santos-Buch & Teixeira 1974, Ribeiro dos Santos & Hudson 1980, Kierszenbaum 1985), a theory that hampered efforts to develop chemotherapy and vaccine against T. cruzi (Kierszenbaum 2005). The importance of parasite persistence in CCC pathogenesis was finally reaffirmed nearly 80 years after Chagas’ pioneering work (Chagas 1916) using immunohistochemistry and molecular biology to analyse the heart tissue of CCC patients (D’Avila Reis et al. 1993, Higuchi et al. 1993, Jones et al. 1993, Reis et al. 1997), as well as ingenious cell transfer studies in experimental models (Tarleton et al. 1997). Currently, it is widely believed that chagasic cardiac injury results from an imbalanced effector immune response elicited by parasite persistence (Higuchi et al. 2003, Kierszenbaum 2005).

Adopting the experimental model consisting of female C3H/He mice infected with low-dose inocula of the Colombian T. cruzi strain, we demonstrated the presence of parasite nests and the persistence of parasite DNA in inflammatory cardiac lesions (Fig. 1), reproducing earlier findings in chronic chagasic patients (Higuchi et al. 1993, Jones et al. 1993). Our model also replicates Chagas’ main findings in CCC patients (Chagas 1916, Chagas & Vilella 1922), including (i) parasitaemia control and acute phase survival; (ii) chronic increase in relative heart weight; (iii) acute and chronic myocarditis; (iv) cardiomyocyte lesions and (v) electrical abnormalities (Fig. 2).

In the past several years, many groups have sought to elucidate the physiopathogenesis of CCC and cardiomyopathy (Higuchi et al. 2003, Marin-Neto et al. 2008). Nevertheless, new therapeutic approaches, beyond already available anti-parasite drugs and interventions targeting symptoms, have not been developed (Marin-Neto et al. 2008, Soeiro & de Castro 2009). Since cardiac transplant is currently the only effective treatment for Chagas disease-induced heart failure, cell-based therapy aiming to recover heart function is a desirable alternative, although expensive and still restricted to clinical trials (Soares & Ribeiro dos Santos 2009). The major challenge in designing efficacious treatment for T. cruzi-elicited cardiomyopathy is to reduce inflammation,
which causes severe tissue damage, without hampering immunological control of parasitism. A more thorough understanding of CCC pathogenesis and its underlying molecular mechanisms may thus help to delineate new therapeutic strategies.

Here, we critically review our and other recent data exploring the participation of cell adhesion molecules (CAM) and CC-chemokines and their receptors in the multi-step process of cell migration during T. cruzi-elicited myocarditis and CCC formation. We also offer evidence that these molecular pathways are rational therapeutic targets for modulating detrimental cardiac inflammation during chagasic infection without hampering parasite control.

**Key players in leukocyte migration to injured tissues**

The three crucial steps of leukocyte migration to injured non-lymphoid tissues (rolling, adhesion and transmigration) were first described in the XIX century (Ley et al. 2007). However, only in the last decade of the XX century, after the discovery of molecules involved in the leukocyte migration cascade, has this multi-step...
model emerged as a concept and began to unravel the leukocyte recruitment process to injured tissue (Butcher 1991, Springer 1995). In the original model, Butcher and Springer proposed three steps underlying leukocyte migration: (i) selectin-mediated rolling; (ii) chemokine-driven activation and (iii) integrin-dependent arrest or firm adhesion. However, recent findings have revealed that the leukocyte migration cascade is more complex, involving tethering or capture, rolling, arrest or firm adhesion, recruitment, transendothelial migration (intraluminal crawling and paracellular and transcellular migration), extravasation through the basement membrane and positioning (Ley et al. 2007, Sigmundsdottir & Butcher 2008). Notably, all of these studies confirmed the participation of chemokines, chemokine receptors and CAM in leukocyte trafficking to injured tissues.

Chemokines are small (8-14 kDa) chemoattractant cytokines that are either constitutively expressed or induced in response to inflammation. These proteins comprise four subfamilies (CXC or α, CC or β, C or γ and CX,C or δ) that act on G-protein-coupled serpentine receptors on target cells. Generally, CXC-chemokines act on neutrophils and on T and B cells, while CC-chemokines affect a wider spectrum of cell populations, including monocytes, basophils, eosinophils, T cells, dendritic cells and natural killer cells, but do not act on neutrophils. It is likely that all cell types, such as endothelial cells, cardiomyocytes, fibroblasts, T cells and macrophages, produce a variety of chemokines when adequately stimulated (Bromley et al. 2008, Viola & Luster 2008). Chemokines act in a context-dependent manner, varying in function according to whether they are in soluble or extracellular matrix-anchored form (Sigmundsdottir & Butcher 2008). Their biological functions are numerous, including facilitating (i) cell adhesion to the endothelium and extracellular matrix components; (ii) leukocyte chemotactic migration; (iii) angiogenesis control; (iv) extracellular matrix production control; (v) regulation of T-lymphocyte growth, activation and differentiation (including Th1/Th2 commitment); (vi) release of intracellular reserve of enzymes; (vii) oxygen radical formation; (viii) cytoskeleton alteration; (ix) generation of lipid mediators of intracellular activation signals; (x) haematopoietic precursor proliferation and (xi) control of pathogen invasion and propagation. Chemokines may thus play an important role in the regulation of host resistance to pathogens and in the immunopathogenesis of various conditions, including autoimmune and infectious diseases (Proudfoot 2002, Bromley et al. 2008, Viola & Luster 2008).

CAM, which are mainly members of the selectin, integrin and immunoglobulin superfamilies, participate in all cell migration steps involving cell-cell and cell-extra-cellular matrix interactions (Luo et al. 2007). These molecules are widespread, present on a variety of cell types and involved in diverse biological events, including (i) ontogenesis; (ii) immunological synapse formation; (iii) cell activation; (iv) immunological surveillance; (v) cell extravasation to injured tissue; (vi) cell positioning and (vii) cell-pathogen interactions (Luo et al. 2007, Abram & Lowell 2009). Importantly, chemokine-triggered mod-
ings, it is reasonable to propose that CAM contribute to chagasic myocarditis formation by mediating lymphocyte adhesion to the cytokine-activated endothelium of cardiac blood vessels. Moreover, one could speculate that LFA-1+ lymphocytes may adhere to ICAM-1-bear-
ing cardiomyocytes, contributing to parasite control but also causing tissue lesions (Benvenuti et al. 2000). CAM may thus play an integral role in creating an appropriate scenario for chagasic heart failure, a role that is currently being elucidated experimentally.

Adopting C3H/He mice infected with the Colombi-
an T. cruzi strain, which develop CD8+ T cell-enriched cardiomypathy and increased heart relative weight, cardiomyocyte lesions and ECG abnormalities (Fig. 2), we demonstrated that both CD4+ and CD8+ T cells in-
filtrating the heart tissue exhibit the activation pheno-
type CD62Llow, LFA-1high and VLA-4high. In addition, a higher proportion of circulating CD8+ T cells than CD4+ T cells are LFA-1high and VLA-4high, which may allow interaction of these circulating cells with ICAM-1- and VCAM-1-activated endothelium (dos Santos et al. 2001). This potential interaction could facilitate the preferential migration to and predominance of these T cells in the myocardium of chronically T. cruzi-infected animals. Interestingly, in T. cruzi-infected C3H/He mice, the in-
creased frequency of LFA-1+ and VLA-4+ circulating T cells and the upregulation of ICAM-1 and VCAM-1 on the cardiac vascular endothelium coincide with inflamm-
atory cell invasion of this tissue (Marino et al. 2003). Using interleukin-12 (IL-12) and IFN-γ-deficient mice, we have also found that IFN-γ modulates ICAM-1 ex-
pression on endothelial cells and on the sarcolemma of cardiomyocytes (Michailowsky et al. 2004). However, a study implementing IFN-γ receptor-deficient mice claimed that cardiac ICAM-1 expression in T. cruzi-in-
fected mice is IFN-γ-independent (Laucella et al. 1996b), suggesting that other inflammatory factors or parasite products per se may also contribute to CAM upregula-
tion during T. cruzi infection. In fact, in vitro T. cruzi in-
fection of endothelial cells induces VCAM-1 and E-
selectin and upregulates ICAM-1 via nuclear factor-kB (NF-kB) activation (Huang et al. 1999).

Infection of ICAM-1- mice indicated a role for ICAM-1 in T. cruzi-elicited myocarditis formation. The high susceptibility of ICAM-1-deficient animals to T. cruzi infection was associated with a decreased num-
ber of CD4+ and CD8+ T cells invading the cardiac tissue, implying that LFA-1/ICAM-1-mediated cell migration to the heart is essential for myocarditis formation and thus for protective anti-T. cruzi responses (Michaikowsky et al. 2004). However, whether different CAM have dis-
tinct roles in protective immunity and in the induction of chronic inflammation in Chagas disease is still unclear. A better understanding of the role of CAM in T. cruzi-
elicted myocarditis and CCC may pave the way to facili-
tate targeting of these molecules to treat Chagas disease. CAM antagonists have already been implemented in the treatment of chronic inflammatory diseases, including rheumatoid arthritis, multiple sclerosis and Crohn’s dis-
ease (Kummer & Ginsberg 2006, Abram & Lowell 2009), indicating the feasibility of this therapeutic approach.

Chemokines and chemokine receptors in T. cruzi-
elicted myocarditis and cardiomyopathy

During T. cruzi infection, cytokines, chemokines and other immune mediators are generated in the af-
ected heart tissue, triggering intense leukocyte migration (Talvani et al. 2000, dos Santos et al. 2001, Marino et al. 2004, Medeiros et al. 2009). CC- or β-chemokines appear to be particularly beneficial to the host during T. cruzi infection. The parasite or its molecules have been found to induce human and murine macrophages as well as murine cardiomyocytes to produce CC-chemokines in vitro (Lima et al. 1997, Villalta et al. 1998, Aliberti et al. 1999, Machado et al. 2000, Talvani et al. 2000). Ad-
dition of macrophage inflammatory protein-1α (MIP-1α, or CCL3), MIP-1β (or CCL4) or RANTES (Regulation on Activation, Normal T-cell Expressed and Secreted or CCL5) to such infected macrophages and cardiomyo-
cytes resulted in increased parasite uptake and destruc-
tion by a nitric oxide-dependent mechanism (Aliberti et al. 1999, Machado et al. 2000). This finding suggests that parasites regulate chemokine production, which keeps the parasite under control and thus prevents host death.

Treatment of T. cruzi-infected cardiomyocytes with TNF also leads to induction or augmentation of CC-
chemokine expression (Machado et al. 2000). Interest-
ingly, TNF- or IFN-γ-treated T. cruzi-infected macrophages express more CC-chemokines than untreated cells, while IL-10 and transforming growth factor-β (TGF-β) reduce CC-chemokine production, suggesting that pro-inflam-
matory and regulatory cytokines control parasitism and myocarditis via chemokine-dependent mechanisms (Aliberti et al. 1999). Additionally, the CC-chemokine monocyte chemotactant protein-1 (MCP-1 or CCL2) directly triggers the chemotaxis and morphogenesis of the trypomastigote forms of T. cruzi (Yamauchi et al. 2006), implicating CC-chemokines as crucial actors in directing parasite to certain tissues and contributing to explain T. cruzi tissue tropism (Melo & Brener 1978).

Elevated mRNA levels of several chemokines [CCL2/ MCP-1, CCL3/MIP1α, CCL4/MIP-1β, CCL5/RANTES, monokine induced by IFN-γ (MIG or CXCL9), interfer-
on-inducible protein-10 (IP-10 or CXCL10) and RANT-
es] have been observed in the cardiac tissue, associated with the formation of inflammatory infiltrates of various murine models of acute and chronic T. cruzi infection (Talvani et al. 2000, Aliberti et al. 2001, dos Santos et al. 2001, Marino et al. 2004). Indeed, we recently demonstrated that myocarditis intensity during both the acute and the chronic phases of infection in C3H/He mice is directly correlated with the CC-chemokines CCL3/MIP-
1α and CCL5/RANTES but not the cytokines IFN-γ and TNF, concentrations in the cardiac milieu (Medeiros et al. 2009), reaffirming CC-chemokine participation in T. cruzi-elicited myocarditis formation and perpetuation.

Treatment of Colombian T. cruzi-infected BALB/c mice with antibodies recognizing CXCL10/IP-10 or CCL5/RANTES during the acute phase did not alter parasitaemia, heart parasitism or inflammation. How-
ever, anti-CXCL9/Mig (monokine induced by IFN-γ) associated with anti-CXCL10/IP-10 led to increased
parasitaemia and parasitism but did not affect either myocarditis severity or the transcript levels of inflammatory cell markers (CD4, CD8, F4/80), cytokines (TNF or IFN-γ) and chemokines (CXCL9/MIG, CXCL10/IP-10, or CCL5/RANTES) in the heart tissue, as compared with isotype-treated control mice (Hardison et al. 2006c). These data indicate that CXC-chemokines promote a protective immune response to parasite without contributing to cardiac inflammation.

The role played by CC-chemokines in myocarditis and cardiomyopathy development during T. cruzi infection has also been explored. Treatment of acutely infected mice with neutralising antibodies revealed that CCL3/MIP-1α is involved in macrophage recruitment to infected tissue (Petray et al. 2002). Moreover, CCL3/MIP-1α-deficient mice infected with the Brazil T. cruzi strain contrasting with wild-type mice presented normal cardiac activity, suggesting that CCL3/MIP-1α plays an important role in cardiac remodelling during infection (Durand et al. 2006). Our recent data indicate that CCL3/MIP-1α controls macrophage and CD8+ T-cell, but not CD4+ T-cell, migration to the heart tissue, particularly during the chronic phase. In addition, CCL3/MIP-1α influences CD8+CCR5+ T-cell compartmentalisation, which may explain the reduced migration of these cells into the cardiac tissue. Our data also indicate that CCL3/MIP-1α controls cardiac parasitism and cardiomyocyte lesion formation (Silverio et al. 2009). Meanwhile, CCL4/MIP-1β may prevent excessive inflammation and T. cruzi pathogenesis, as indicated by a recent study showing that immunisation of animals with a CCL4/MIP-1β DNA vaccine led to the production of antibodies that exacerbated cardiac inflammation and fibrosis without affecting parasitism (Roffé et al. 2006).

Elevated plasma chemokine levels have been correlated with worse disease in patients with heart failure (Filippatos & Kardaras 2002). For example, high levels of CCL2/MCP-1, but not CCL3/MIP-1α, are directly correlated with heart dysfunction in CCC (Talvani et al. 2004a). Since CCC progression is probably multifactorial, involving an interplay between host genetics, the parasite, and environmental factors, characterisation of host genetic predisposition may help to identify risk factors for disease progression. For example, a study of a CCL2/MCP-1 single nucleotide polymorphism at the −2518A/G position revealed that subjects with the CCL2/MCP-1-2518AA genotype have a 4-fold greater risk of developing CCC than those without this genotype (Ramasastry et al. 2006). Although the authors proposed that the CCL2/MCP-1 variant A, correlated with a low transcriptional level, is a genetic modifier of clinical outcome for T. cruzi infection, the CCL2/MCP-1 status of these patients and the participation of CCL2/MCP-1 in CCC progression remain to be clarified. CCL2/MCP-1 signals exclusively through CCR2, which also recognises CCL8/MCP-2, CCL7/MCP-3 and CCL13/MCP-4 (Viola & Luster 2008). CCR2-deficient mice infected with the Colombian T. cruzi strain exhibit a transient increase in heart parasitism but not parasitaemia during acute infection. Neither the intensity of their cardiac inflammation nor their IFN-γ and inducible nitric oxide synthase mRNA levels differs from those of wild-type mice (Hardison et al. 2006a). Conversely, CCL2−/− mice infected with the Y strain of T. cruzi are more susceptible to infection, presenting higher parasitaemia and parasitism.

In CCL2/MCP-1-deficient mice, macrophages display reduced trypanocidal activity and CCL2/MCP-1 has been shown to be important for macrophage and CD8+ T-cell activation, cell migration to the cardiac tissue and inflammatory cell retention in focal infiltrates (CN Paiva et al., unpublished observations). Myocarditis intensity in acutely and chronically Colombian strain-infected C3H/He mice is thus directly correlated with the cardiac concentration of the CC-chemokine CCL2/MCP-1 (our unpublished observations). Together, these data indicate that the effects of CC-chemokines on the differential migration of distinct cell populations into the cardiac tissue and effector activity of the infiltrating inflammatory cells during T. cruzi infection should be further explored.

Chemokine receptor binding leads to integrin activation, encouraging firm leukocyte adhesion to the activated endothelium and extravasation into the inflamed tissues (Viola & Luster 2008). Thus, characterisation of chemokine receptor expression on circulating leukocytes may foster better comprehension of the roles of different cell populations in tissue injury. Peripheral blood mononuclear cells (PBMCs) of chagasic patients tend to express CCR5 and CXCR4 at a higher frequency than those of non-infected individuals (Talvani et al. 2004b). A high frequency of circulating CCR5+ lymphocytes, which were mainly CD8+, was also detected in acutely T. cruzi-infected mice (Marino et al. 2004). Interestingly, this increase in CCR5+ cells among the PBMC population occurs 14 days post-infection (Silva et al. 2007), coinciding with augmentation of LFA-1highVAL4high activated PBMC numbers and myocardium colonisation by inflammatory cells (Marino et al. 2003). Studies of CCR5 59029A/G gene polymorphism in Peruvian (Calzada et al. 2001) and Venezuelan patients (Ferrandez-Mestre et al. 2004) revealed that the G allele, which reduces CCR5 expression, is found more frequently in asymptomatic patients than in cardiopaths. Together, these data suggest that CCR5 may be linked to CCC severity. In vitro stimulation of PBMCs from CCC patients with T. cruzi antigens also enhances the frequency of CD4+ and CD8+ T lymphocytes co-expressing CCR5/IFN-γ, CXCR3/IFN-γ and CXCR3/TNF (Gomes et al. 2005). Meanwhile, asymptomatic patients had an increased frequency of CD4+ T and CD8+ lymphocytes co-expressing CCR3/IL-10 and CCR3/IL-4. Therefore, CCC genesis may be correlated with the migration of cells bearing chemokine receptors and expressing specific effector functions from the peripheral blood to the cardiac tissue (Lannes-Vieira 2003). In the cardiac tissue of Colombian strain-infected C3H/He mice, there is a predominance of CD8+ CCR5+ T lymphocytes relative to CCR5+CD4+ T cells (Marino et al. 2004). To modulate heart inflammation, mice were treated with N-terminal-methionylated RANTES (Met-RANTES), a selective partial antagonist that acts on CCR1 and CCR5. This drug did not interfere with heart parasitism but significantly reduced the
numbers of CD4+; CD8+; and CCR5+ lymphocytes in the tissue. Furthermore, decreased fibronectin deposition paralleled an increase in the survival of treated animals. Notably, the number of IFN-γ+ cells in the heart was similar in saline-injected and Met-RANTES-treated mice, while the IL-4+ cell count was drastically reduced in the treated mice (Marino et al. 2004). The cardiac tissue of the Met-RANTES-treated mice also exhibited lower TNF and higher IL-10 levels (Fig. 3), reflecting a more balanced cytokine milieu. These data strongly suggest that while CCR5+ cells are not crucial for parasite control, they may play an important role in the pathogenesis of chagasic cardiomyopathy. Importantly, Met-RANTES administration also demonstrated that T. cruzi-elicited acute meningoencephalitis is a CCR5/CCR1-independent process (Silva et al. 2007), supporting CCR5 targeting as a safe potential therapeutic strategy for modulating chagasic myocarditis.

The predominance of CD8+CCR5+ cells in the heart tissue of T. cruzi-infected animals was corroborated using C57BL/6 mice infected with the Y T. cruzi strain (Machado et al. 2005), indicating that CD8+CCR5+ cell prevalence results from T. cruzi infection and is not an artefact of a particular model. Increased CCR5 expression by CD8+ T cells was detected after in vitro stimulation with a T. cruzi antigen (Machado et al. 2005), demonstrating the central role of the parasite in CCR5 modulation. We have also recently demonstrated that TNF signalling via TNFR1 is crucial for CD8+CCR5+ differentiation, CD8+CD44+CD62Llow compartmentalisation and CD8+ T cell-enriched myocarditis formation during T. cruzi infection (Kroll-Palhares et al. 2008).

CCR5-deficient mice present an intense decrease in macrophage and T-cell migration to the heart, coupled with high parasitaemia and cardiac parasitism, especially during early infection (Hardison et al. 2006b). We recently confirmed that CCR5 is not requisite to the acute-phase survival of Colombian strain-infected mice (NF Vinagre et al., unpublished observations), supporting our proposal that CCR5 may be a rational target for immunoregulation of T. cruzi infection.

The intensity of chronic T. cruzi-elicited myocarditis is related to the concentrations of the CC-chemokines CCL3/MIP-1α and CCL5/RANTES in the heart tissue and to the frequency of peripheral CCR5LFA-1+ T lymphocytes (Medeiros et al. 2009). Given the lack of therapeutic prospects for chronic T. cruzi-elicited cardiac inflammation and our promising results using Met-RANTES to treat acute infection (Marino et al. 2004), we administered this partial CCR1/CCR5 antagonist to chronically Colombian strain-infected mice. This treatment led to a 20-30% decrease in CD4+ cell numbers, reduced IL-10, IL-13 and TNF expression and impaired re-compartmentalisation of activated CD4+CCR5+ lymphocytes. Notably, a beneficial effect of CCR1/CCR5 blockage during chronic infection was revealed by protection against connexin 43 loss in the heart tissue and creatine kinase cardiac isoenzyme MB (CK-MB) level enhancement, both of which are typically markers of heart dysfunction (Medeiros et al. 2009).

Trafficking of immunocompetent CD8+ T cells to the cardiac tissue contributes to chronic T. cruzi-elicited myocarditis in chagasic patients (Reis et al. 1993, Higuchi et al. 1997) and experimental models (dos Santos et al. 2001, Medeiros et al. 2009). There is also a correlation between CD8+ T cell numbers and parasite persistence in the heart tissue of chagasic patients (Reis et al. 1997). Emerging evidence suggests that CD8+ effector-cell sub-populations with varied cytotoxic and pro-inflammatory (IFN-γ) abilities may determine the outcome of an inflammatory process (Sandberg et al. 2001). For instance, there is a sensible correlation between the numbers of IFN-γ+ and TNF+ cells and CD8+ T cells and successful parasite control in CCC patients (Reis et al. 1997). In contrast, the presence of heart-infiltrating, granzyme A-expressing CD8+ cytotoxic T lymphocytes is associated with severe CCC (Reis et al. 1993). In this vein, our recent data showed that the number of perforin+ and IFN-γ+ cells invading the cardiac tissue of T. cruzi-infected....
ed mice was associated with CK-MB release and electrical abnormalities as infection progressed. Although the perforin-dependent pathway played a role in anti-T. cruzi immunity and acute-phase survival, it was not essential when mice were infected with a low inoculum of the Colombian strain. Moreover, perforin deficiency resulted in lower CK-MB isoenzyme activity in the serum and restricted connexin 43 loss, both markers of cardiac lesions and hampered the development of electrical abnormalities. The numbers of IFN-γ-producing cells were also enhanced in perforin-deficient animals, suggesting that this cell population may play a protective role in T. cruzi-induced cardiomyopathy (Silverio et al. 2009).

The data discussed above support the idea that an appropriately intense and focussed immune response can control T. cruzi dissemination in the absence of disease, as observed in patients presenting an indeterminate form of Chagas disease. Hence, a thorough understanding of the biological forces governing leukocyte trafficking to T. cruzi-infected heart tissue, such as cytotoxic or inflammatory CD8+ T cells, may facilitate the design of treatments that regulate CCC without hampering parasite control. Currently, therapies using antibodies to and antagonists of CAM and chemokine receptors are being developed to selectively curb chronic inflammation in degenerative diseases (Kummer & Ginsberg 2006, Ley et al. 2007, Viola & Luster 2008, Horuk 2009). Therefore, the application of these strategies in parasite-triggered chronic inflammation, such as Chagas disease, to regain homeostatic balance in parasite-host interactions is an increasingly rational, attractive and realistic possibility.

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REFERENCES


Butcher EC 1991. Leukocyte-endothelial cell recognition: three (or more) steps to specificity and diversity. Cell 67: 1033-1036.


Hardison JL, Kuziel WA, Manning JE, Lane TE 2006a. Chemokine CC receptor 2 is important for acute control of cardiac parasitism but does not contribute to cardiac inflammation after infection with Trypanosoma cruzi. J Infect Dis 193: 1584-1588.

Hardison JL, Wrightsman RA, Carpenter PM, Kuziel WA, Lane TE, Manning JE 2006b. The CC chemokine receptor 5 is important in control of parasite replication and acute cardiac inflammation following infection with Trypanosoma cruzi. Infect Immun 74: 135-143.
2006c. The chemokines CXCL9 and CXCL10 promote a protective immune response but do not contribute to cardiac inflammation following infection with Trypanosoma cruzi. Infect Immun 74: 125-134.


