Molecular characterisation of Bartonella species in cats from São Luís, state of Maranhão, north-eastern Brazil

Maria do Socorro Costa de Oliveira Braga1,2, Pedro Paulo Vissotto de Paiva Diniz1, Marcos Rogério André1, Caroline Plácidi de Bortoli1, Rosangela Zacarias Machado1/+ 

1Departamento de Patologia Veterinária, Faculdade de Ciências Agrárias e Veterinárias, Universidade Estadual Paulista, Jaboticabal, SP, Brasil 2Universidade Estadual do Maranhão, São Luís, MA, Brasil 3College of Veterinary Medicine, Western University of Health Sciences, Pomona, CA, USA

Bartonella species are fastidious bacteria that predominantly infect mammalian erythrocytes and endothelial cells and cause long-lasting bacteraemia in their reservoir hosts. Reports that describe the epidemiology of bartonellosis in Brazil are limited. This study aimed to detect and characterise Bartonella spp DNA from cat blood samples in São Luís, Maranhão, north-eastern Brazil. Among 200 cats tested for multiple genes, nine (4.5%) were positive for Bartonella spp: six cats for Bartonella henselae and three for Bartonella clarridgeiae. Based on the phylogenetic analysis of four genes, the B. henselae strain matched strains previously observed in Brazil and was positioned in the same clade as B. henselae isolates from the United States of America. Moreover, sequence alignment demonstrated that the B. clarridgeiae strain detected in the present study was the same as the one recently detected in cats from southern Brazil.

Key words: Bartonella henselae - Bartonella clarridgeiae - cats - north-eastern Brazil

Bartonella species are fastidious Gram-negative bacteria that predominantly infect mammalian erythrocytes and endothelial cells and cause long-lasting bacteraemia in their reservoir hosts. Bartonella spp DNA has been molecularly detected in wild felids in Brazil (Filoni et al. 2006) and free-ranging wild felids in Brazil (Filoni et al. 2006) and Rio de Janeiro (RJ) (Souza et al. 2008). There are few studies addressing the occurrence of Bartonella spp in humans in Brazil. Antibodies to B. henselae and Bartonella quintana have been detected in 13.7% and 12.8% of healthy residents of the state of Minas Gerais (MG), respectively (da Costa et al. 2005). A seroprevalence of Bartonella spp of 38.4% and 34.4% has been observed in HIV-positive patients and healthy human populations, respectively, in Jacarepaguá, RJ (Lamas et al. 2010). This study aimed to detect and characterise Bartonella spp DNA from Brazilian cat blood samples from São Luís, Maranhão (MA), north-eastern Brazil.

**MATERIALS AND METHODS**

**Sample collection and study area** - Between October 2008-January 2009, whole blood samples were collected from 200 domestic cats (102 males, 98 females) that were allowed outside in São Luís (194 cats from suburban areas of São Luís county and six from Raposa county). The study region is an invasion area that has suffered a 70% forest reduction. Cats were selected without specific inclusion criteria and were apparently healthy at the time of sample collection. Sampled cats had contact with other cats and dogs. Blood samples for polymerase chain reaction (PCR) were collected from jugular or cephalic veins, immediately aliquoted into tubes containing ethylenediamine tetraacetic acid (EDTA) as an anticoagulant and stored at -20°C until PCR analysis. All procedures were performed according to Ethical Principles in Animal Research adopted by the Brazilian College of Animal Experimentation and were approved by the Ethical Committee of the College of Agricultural Sciences and Veterinary, São Paulo State University (UNESP) Jaboticabal, state of São Paulo (SP) (2008/025862-07).

Financial support: CNPq (#479162/2007-7), WesternU + Corresponding author: zacarias@fcav.unesp.br
Received 21 November 2011
Accepted 14 March 2012

online | memorias.ioc.fiocruz.br
**PCR - DNA was extracted from 200 µL of EDTA-anticoagulated blood with the QIAamp DNA Blood Mini kit** (Qiagen, Valencia, CA, USA) according to the manufacturer’s instructions. The DNA was eluted in 200 µL of EB Buffer (supplied by the DNA extraction kit).

* Bartonella* genus screening was performed by PCR in which the intergenic transcribed spacer (ITS) was targeted, as described previously (Diniz et al. 2007a). Amplification was performed by conventional PCR in a 25 µL final reaction volume that contained 1X PCR mix (Premix Ex Taq, Takara Bio Inc, Shiga, Japan), 12.5 pmol each primer (Integrated DNA Technologies Inc, Coralville, IA) and 5.0 µL DNA template. PCR was performed in a thermocycler (Mastercycler Pro Gradient S, Eppendorf, Hamburg, Germany) under the following conditions: a single hot-start cycle at 95ºC for 2 min, followed by 55 cycles of denaturing at 94ºC for 15 s, annealing at 66ºC for 20 s and extension at 72ºC for 20 s. Amplification was performed by conventional PCR in a thermocycler (Mastercycler Pro Gradient S, Eppendorf, Hamburg, Germany) under the following conditions: a single hot-start cycle at 95ºC for 2 min, followed by 55 cycles of denaturing at 94ºC for 15 s, annealing at 66ºC for 20 s and extension at 72ºC for 20 s. Amplification was completed by an additional cycle at 72ºC for 1 min and products were resolved in a 1.5% agarose matrix by electrophoresis and analysed under ultraviolet exposure. DNA from a healthy, pathogen-free cat was used as a PCR negative control. DNA extracted from *B. quintana* (similar to GenBank accession L35100) was used as a positive control. To prevent PCR contamination, the sample extraction, reaction setup, PCR amplification and amplicon detection were performed in separate areas.

Additionally, when genomic material was available, samples were evaluated for the presence of five additional genes: the bacteriophage-associated heme-binding protein gene (*pap31*) (Diniz et al. 2007a), the RNA polymerase beta subunit gene (*rpoB*) (Diniz et al. 2007a), the riboflavin synthase gene (*ribC*) (Johnson et al. 2003), the heat shock protein gene (*groEL*) (Barber et al. 2010) and the citrate synthase gene (*gltA*) (Winoto et al. 2005). The amplification conditions were similar to those used for the ITS, but different annealing temperatures were used for each primer pair: 62ºC for *pap31* and *rpoB*, 55ºC for *ribC* and *gltA* and 58ºC for *groEL*.

**Phylogenetic analysis -** Amplicons were gel-purified or PCR-purified (MiniElute kit, Qiagen, Valencia, CA, USA) and sequenced by the Department of Technology - UNESP or by Eurofins MWG Operon (Huntsville, AL, USA). Chromatogram evaluations, primer deletions and sequence alignments were performed with the software Contig-Express and AlignX (Vector NTI Suite 10.1, Invitrogen Corp, Carlsbad, CA, USA). Bacterial species and strain types were defined by comparison with other sequences deposited in the GenBank database with the Basic Local Alignment Search Tool (Altschul et al. 1990). Phylogenetic analysis was performed with concatenated sequences of the ITS, *rpoB*, *ribC* and *gltA* genes with the maximum likelihood method based on the Kimura two-parameter model (Kimura 1980) with MEGA4 software (Tamura et al. 2007). The *pap31* and *groEL* sequences were excluded from the concatenated phylogenetic analysis because the available DNA sequences in GenBank from other *Bartonella* species were limited.

**RESULTS**

Among 200 cats tested for multiple genes, nine (4.5%) were positive for *Bartonella* spp: six for *B. henselae* (4 males, 2 females) and three for *B. clarridgeiae* (1 male, 2 females) (Table). These isolates were positioned in the same clade as other *B. henselae* isolates from dogs and humans from Brazil and the United States of America (USA) and the *B. clarridgeiae* isolate from a cat from the USA, with high bootstrap support (100/100). Sixty (30%) of the cats were parasitized by fleas (*Ctenocephalides felis*), including cats positive for *Bartonella* spp.

**TABLE**

Species of *Bartonella* identified in five cats by DNA amplification and sequencing from São Luis, state of Maranhão, Brazil

<table>
<thead>
<tr>
<th>Cat number</th>
<th>ITS</th>
<th>rpoB</th>
<th><em>pap31</em></th>
<th><em>ribC</em></th>
<th><em>gltA</em></th>
<th><em>groEL</em></th>
</tr>
</thead>
<tbody>
<tr>
<td>3</td>
<td>B.h.</td>
<td>B.h.</td>
<td>B.h.</td>
<td>B.h.</td>
<td>B.h.</td>
<td>NT</td>
</tr>
<tr>
<td>6</td>
<td>B.h.</td>
<td>B.h.</td>
<td>B.h.</td>
<td>B.h.</td>
<td>B.h.</td>
<td>NT</td>
</tr>
<tr>
<td>7</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>B.c.</td>
<td>-</td>
<td>B.c.</td>
</tr>
<tr>
<td>8</td>
<td>B.c.</td>
<td>-</td>
<td>-</td>
<td>B.c.</td>
<td>NT</td>
<td>NT</td>
</tr>
<tr>
<td>10</td>
<td>B.h.</td>
<td>B.h.</td>
<td>B.h.</td>
<td>B.h.</td>
<td>B.h.</td>
<td>B.h.</td>
</tr>
<tr>
<td>109</td>
<td>B.h.</td>
<td>-</td>
<td>B.h.</td>
<td>B.h.</td>
<td>B.h.</td>
<td>NT</td>
</tr>
<tr>
<td>120</td>
<td>B.h.</td>
<td>-</td>
<td>B.h.</td>
<td>B.h.</td>
<td>B.h.</td>
<td>NT</td>
</tr>
<tr>
<td>134</td>
<td>B.h.</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>B.h.</td>
<td>NT</td>
</tr>
<tr>
<td>187</td>
<td>B.c.</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>NT</td>
</tr>
</tbody>
</table>

We deposited several partial *B. henselae* Brazil-1 sequences into the GenBank database, including HQ012581 (ITS), HQ012584 (rpoB), HQ012582 (pap31), HQ012583 (ribC), HQ012580 (gltA) and HQ074721 (groEL). Additionally, several *B. clarridgeiae* sequences were deposited, including HQ012586 (rpoB), HQ012585 (ribC) and HQ074720 (groEL).

**DISCUSSION**

The ITS and rpoB sequences obtained from three cats infected with *B. henselae* were 100% homologous (ITS = 546/546 bp, rpoB = 585/585 bp) to sequences of the *B. henselae* Brazil-1 strains previously detected from a 36-year-old, HIV antibody-positive Brazilian male with severe anaemia, pancreatitis, ascitis and cryptogenic hepatitis (Velho et al. 2007) and from a dog in Brazil with fever, severe anaemia, thrombocytopenia and myocarditis (Diniz et al. 2007a). In addition, the partial *pap31* sequence obtained from these three cats was 100% homologous (490/490 bp) to the *B. henselae* Brazil-1 strain detected from the sick dog (Diniz et al. 2007a), but not compared to the human case because *pap31* sequences were not obtained in that reported case (Velho et al. 2007). This is the first description of this *B. henselae* strain infecting cats in Brazil and these results may represent an important aspect of the epidemiology of this organism. The entire genome sequence of the *B. henselae* Brazil-1 strain is not available, but based on the phylogenetic analysis of the four genes (Figure), this strain is positioned in the same clade as other *B. henselae* isolates. Because the cats evaluated in this study are located in a distinct region of Brazil, approximately 2.880 km (1.789 miles) from the region in which the human and canine cases were originally reported, the *B. henselae* Brazil-1 strain may represent the endogenous *B. henselae* strain of Brazil. A recent study in the Southern region of Brazil detected two *B. henselae*-infected cats with ribC sequences similar to the Brazil-1 strain (HM588661 and HM588662); however, other genes were not amplified and sequenced (Staggemeier et al. 2010). Our results expand the molecular characterisation of the *B. henselae* Brazil-1 strain, with partial ribC (536 bp), gltA (915 bp) and groEL (565 bp) gene sequences that were not available for the human or canine cases previously reported. These new DNA sequences provide important data for future multilocus sequence analysis and phylogenetic studies of *B. henselae* strains.

Three cats were infected with *B. clarridgeiae* and the sequences of the rpoB, ribC and groEL genes from two of these cats revealed that this species was 100% homologous to a strain of *B. clarridgeiae* (ATCC 51734) that was isolated from a kitten in Texas, USA (Clarridge et al. 1995). Moreover, it was also similar to a strain of *B. clarridgeiae* (CIP 104772) that was isolated from a cat in Strasbourg, France (Lawson & Collins 1996). Multiple sequence alignments of the ribC gene demonstrated that these two cats from the north-eastern region of Brazil were infected with the same strain recently detected from one of three cats infected with *B. clarridgeiae* in the southern region of Brazil (HM588660) (Staggemeier et al. 2010). In addition, unpublished DNA sequences deposited in GenBank indicate that the same sequence was identified in China (EU571943 and EU836705), although the host species was not indicated.

Fleas are responsible for the transmission of *B. henselae* and *B. clarridgeiae* among cats (Chomel et al. 1996, Foil et al. 1998). Thirty percent of the cats sampled in this study were infested with *C. felis* (including those positive for *Bartonella* spp by PCR), but were apparently healthy at the moment of blood collection. An increasing number of arthropod vectors, including keds, lice, biting flies, sandflies and ticks, have been observed or suspected to be associated with the transmission of *Bartonella* spp among animal populations (Breitschwerdt et al. 2010a). Although numerous molecular surveys have detected *Bartonella* DNA in ticks, predominantly in dogs and humans, there is little evidence that *Bartonella* spp can replicate within ticks and no definitive evidence of transmission by a tick to a vertebrate host (Angelakis et al. 2010). Although vertical transmission of *Bartonella* spp among natural rodent hosts has been confirmed (Kosoy et al. 1998), *B. henselae* is not transmitted transplacentally, via colostrum or milk, or venereally, among cats.
Bartonella spp in cats from Maranhão • Maria do Socorro Costa de Oliveira Braga et al. 775
cats (Abbott et al. 1997, Guptill et al. 1998). Blood trans-
fusion is another route of Bartonella spp transmission
among cats (Kordick et al. 1999).

Cats may be more likely to exhibit clinical symp-
toms upon infection with a non-reservoir-adapted Bar-
tonella species (Breitschwerdt et al. 2010a). Bartonella
spp induces asymptomatic infection in preferred hosts,
behaving like a stealth pathogen (Kordick & Breitsch-
werdt 1998). However, Bartonella infections in cats have
been associated in some cases with fever, lethargy, an-
orexia, reproductive failure, lymphadenopathy, stomati-
tis, uveitis, endocarditis and neurological dysfunction
(Glaus et al. 1997, Guptill et al. 1998, Lappin & Black
1999, Breitschwerdt et al. 2010a, Dowers et al. 2010).

Several Bartonella species are considered zoonotic,
including B. henselae and B. clarridgeiae. Cats are the
primary reservoir for B. henselae and likely B. clarr-
idgeiae and are the transmission vector for human in-
festation. The most frequent infection route to humans
is through scratches contaminated with flea faeces
(Chomel et al. 1996). Cat bites are a less likely route,
as shedding of B. henselae in cat saliva has not been
clearly documented (Chomel et al. 2006a). Recently, evi-
dence for B. vinsonii subsp. berkhoffii and B. henselae
transmission to children in utero or during caesarean
section has been reported (Breitschwerdt et al. 2010b).

In addition, the ability of B. henselae to adhere and in-
vade mature human erythrocytes (Pitassi et al. 2007)
and remain viable in red blood cells after 35 days of
storage at 4°C (Magalhães et al. 2008) underscore the
possibility that these organisms may be transmitted by
Although immunocompromised patients may present
with a systemic and fatal disease, immunocompetent
individuals infected with Bartonella spp may develop
non-specific and usually non-life-threatening clinical
symptoms that have a significant impact on quality of
life (Breitschwerdt et al. 2008, 2010a, b, Kaiser et al.
2011, Kalogeropoulos et al. 2011). Epidemiologic data on
human bartonellosis in Brazil is very limited. Among
457 healthy adults in the countryside of MG, 13.7% and
12.8% harboured B. henselae and B. quintana (cut-off
titre of 64), respectively (da Costa et al. 2005). Another
study in a semi-rural area of RJ detected Bartonella spp
seroprevalences of 38.4% and 34.4% (cut-off titre of 20)
among 125 asymptomatic HIV-positive patients and 125
blood donors (Lamas et al. 2010). In this study, contact
with cats was identified as a risk factor for Bartonella
exposure. Clinical cases associated with high IgG titres
against Bartonella spp have been reported in Brazil, in-
cluding two fatal cases of culture-negative endocarditis
(Siciliano et al. 2006) and one asymptomatic case (La-
mas et al. 2007). Additionally, a 40-year-old man tested
seropositive for Bartonella spp, likely from a cat scratch,
which was complicated with aseptic meningitis and neu-
roretinitis (Pinto Júnior et al. 2008). Ophthalmic find-
ings, including retinal infiltrates, angiomatous lesions,
neuroretinitis and visual loss, were associated with
general symptoms (fever, lymphadenopathy, spleen and
liver enlargement and rash) in cat scratch disease cases
diagnosed in human patients who were seropositive for
Bartonella spp (Curi et al. 2010).

The percentage of Bartonella spp-positive cats identi-
ified by PCR in the present study was lower than that in
other regions of Brazil. For example, a molecular study
of cats from RS observed that 17% of 47 feral cats were
infected with Bartonella spp; five (10.6%) were infected
with B. henselae and three (6.4%) were infected with B.
clarridgeiae (Staggemeier et al. 2010). Most cats from an
animal shelter in city of Vassouras, RJ were infected
(97.3% PCR-positive) (Souza et al. 2010). Recently, B.
henselae DNA and antibodies to Bartonella spp were de-
tected in 42.5% and 47.5% of cats, respectively, undergo-
ning neutering or spaying in RJ (Cripps & et al. 2011).

Further detection and genetic sequencing of Bartonel-
la spp from multiple areas in Brazil is needed to more
precisely characterise their geographic distribution, preva-
ence, new hosts and reservoirs and zoonotic impact.

ACKNOWLEDGEMENTS
To Dr Ricardo Maggi, for sharing unpublished sequences of Bartonella species for phylogenetic comparison.

REFERENCES
Abbott RC, Chomel BB, Kasten RW, Floyd-Hawkins KA, Kikuchi Y,
Koehler JE, Pedersen NC 1997. Experimental and natural in-
festation with Bartonella henselae in domestic cats. Comp Immunol

Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ 1990. Basic

Angelakis E, Billeter SA, Breitschwerdt EB, Chomel BB, Raoul D
2010. Potential for tick-borne bartonelloses. Emerg Infect Dis
16: 385-391.

Avidor B, Graidy M, Efrat G, Leibowitz C, Shapira G, Schattner A,
Zimhony O, Giladi M 2004. Bartonella koehlerae, a new cat-
associated agent of culture-negative human endocarditis. J Clin
Microbiol 42: 3462-3468.

Barber RM, Li Q, Diniz PP, Porter BF, Breitschwerdt EB, Claiborne
MK, Birkenheuer AJ, Levine JM, Levine GJ, Chandler K, Ken-
ny P, Nghiem P, Wei S, Greene CE, Kent M, Plat7 SR, Greer K,
Schatzberg SJ 2010. Evaluation of brain tissue or cerebrospinal
fluid with broadly reactive polymerase chain reaction for Ehrli-
chia, Anaplasma, spotted fever group Rickettsia, Bartonella and
Borrelia species in canine neurological diseases (109 cases). J Vet

Breitschwerdt EB, Maggi RG, Chomel BB, Lappin MR 2010a. Bar-
tonellosis: an emerging infectious disease of zoonotic import-
ance to animals and human beings. J Vet Emerg Crit Care (San An-
tonio) 20: 8-30.

Breitschwerdt EB, Maggi RG, Farmer P, Mascarelli PE 2010b. Mo-
lecular evidence of perinatal transmission of Bartonella vinso-
nii subsp. berkhoffii and Bartonella henselae to a child. J Clin
Microbiol 8: 2289-2293.

Breitschwerdt EB, Maggi RG, Nicholson WL, Cherry NA, Woods
CW 2008. Bartonella sp. bacteremia in patients with neurological

Chomel BB, Boulouis HJ, Breitschwerdt EB 2004. Cat scratch disease
and other zoonotic Bartonella infections. J Am Vet Med Assoc
224: 1270-1279.

Chomel BB, Boulouis HJ, Maruyama S, Breitschwerdt, EB 2006a.
Bartonella spp in pets and effect on human health. Emerg Infect
Dis 12: 389-394.

Chomel BB, Kasten RW, Floyd-Hawkins K, Chi B, Yamamoto K,
Roberts-Wilson J, Furish FN, Abbott RC, Pedersen NC, Koeh-


