Association between *Helicobacter pylori*, *cagA*, and *vacA* Status and Clinical Presentation in Iranian Children

Mandana Rafeey, MD; Reza Ghotaslou*, MD, PhD; Morteza Milani, PhD; Nima Farokhi, MD; Morteza Ghojazadeh, MD, PhD

Tabriz University of Medical Sciences, Tabriz, Iran

Received: Jan 19, 2013; Accepted: Jun 29, 2013; First Online Available: Aug 15, 2013

Abstract

Objective: Seroprevalence of *H. pylori* infection in Iran exceeds 65% of pediatric population. In this study, we intended to find association between the virulence genes (*caqA* and *vacA*) and clinical presentations.

Methods: H. pylori isolates were achieved from the gastric mucosa of children. In each case, the gastric biopsy specimens were cultured and the organisms identified. Detection of different genotypes was carried out by PCR method.

Findings: A total of 106 biopsy specimens were cultured and 33 *H. pylori* isolates obtained. Among these 33 *H. pylori* strains 24 (73%) were *cagA*-positive. Genotypes of *vacA* s1m2, s1m1, s2m2, and s2m1 were 45.5%, 30.3%, 21.2%, and 3%, respectively. Most female patients were infected with genotype s1m2. The *vacA*-m1 strains were significantly more common in patients with nodular gastritis. There were no statistical differences between the *vacA* and *cagA* genotypes and clinical outcomes.

Conclusion: The frequency of *cagA* genotype was high. In this study, nodular gastritis was a common finding and was rather significantly associated with m1 allele of *vacA*.

Iranian Journal of Pediatrics, Volume 23 (Number 5), October 2013, Pages: 551-556

Key Words: Helicobacter Pylori; CagA Protein; VacA Protein; Peptic Ulcer

Introduction

The frequency of *H. pylori* is high in developing countries^[1]. Its seroprevalence in Iran exceeds 65% of pediatric population^[2-3]. A recent study revealed an early colonization/infection of infants with *H. pylori* with a prevalence of 67% at 9 months of age in Northwest and West Iran^[4-5]. The prevalence varies among countries with existing evidence suggesting that the diversity in disease outcome may be recognized by variations in infecting strains^[6-7]. Histological gastritis is essentially universal among *H. pylori* infected

individuals, but only a minority develops a clinically main outcome, such as peptic ulcer disease, lymphoma or gastric cancer^[8]. H. pylori strain-specific factors may influence the pathogenicity of different H. pylori isolates. H. pylori studies have primarily focused on two groups of bacterial virulence factors as the cag pathogenicity island and the vacA[9-10]. The presence of an intact cag pathogenicity island is associated with increased interleukin-8 production and mucosal inflammation^[9]. Overall, the data support the idea that infection with a cagA positive isolate increases the risk but does

Address: Tabriz University of Medical Sciences, Iran

E-mail: rzgottaslo@yahoo.com

© 2013 by Pediatrics Center of Excellence, Children's Medical Center, Tehran University of Medical Sciences, All rights reserved.

^{*} Corresponding Author;

not predict the presence of a clinically significant outcome^[10]. Differences in the vacA generation of signal and middle region allelic types have been identified, and attempts have been made to associate specific vacA genotypes especially s1m1 type with different outcomes, especially with duodenal ulcer disease^[11]. There are several fresh studies inspecting the association between Н. pylori virulence factors and clinical presentations in Iranian adult patients^[12-13]. In this study, we intended to find the association between virulence genes and clinical outcome.

Subjects and Methods

Patient population and endoscopic evaluation:

H. pylori isolates were achieved from the gastric mucosa of children who underwent endoscopy at Pediatric Hospital, Tabriz University of Medical Sciences. Gastric biopsies were obtained during a diagnostic fiberoptic upper endoscopy performed at the discretion of the pediatric gastroenterologist because of the subjects' persistent gastrointestinal symptoms during one year. Disease diagnosis was defined as follows: normal gross appearance, erosions, ulcers, and nodularity. Patients who received H. pylori therapy within 4 weeks prior to the study were excluded. Informed consent was obtained from parents participants, and the protocol was approved by the ethics committee of Tabriz University of Medical Sciences.

Histopathology evaluation:

One biopsy from the antrum and/or fundus was fixed in formalin and processed for pathologic evaluation. Hematoxylin-eosin, and Giemsa stained slides were graded using the visual analog scale of the Sydney classification, which guided analysis of the density of *H. pylori*, and the amounts of neutrophils, mononuclear inflammatory cells, and intestinal metaplasia^[12].

H. pylori culture and genotyping:

In each case, the gastric biopsy specimens were cultured and the organisms identified as *H. pylori* as previously described^[15]. DNA extraction was obtained by CTAB reagent method^[16]. The

extracted DNA was eluted in 50 μ l of 1 × TE buffer {10 mMTris -HCl, 1 mM EDTA (pH 8.0)} and stored at -20°C until used. The *glmM* (*ureC*) gene was used as a positive control for detecting *H. pylori* DNA. The genotypes of *vacA* s-region (s1 or s2) and m-region (m1 or m2), and the presence of *cagA* genes were carried out by PCR as previously described[15].

Statistical analysis:

Variables such as gender, age and the presence of each candidate gene were evaluated. The univariate association between each genotype and the clinical presentations were quantified by the Chi-square or Fisher's exact tests. Independent samples t and one way ANOVA tests were used to compare quantity variables. A *P* value of less than 0.05 was accepted as statistically significant. The SPSS statistical software package version 16.0 was used for all statistical analyses.

Findings

The patients' demographic characteristics, endoscopic presentation, and histological scores are presented in Table 1. A total of 106 specimens of biopsy mucosa were available for histological analysis and the results also are summarized in Table 1. The mean age of patients was $8.28 (\pm 1.59)$ years (rnge: 2-17). After processing for culture only 33 patients had *H. pylori* positive cultures. Most common finding on the basis of endoscopy was nodular gastritis. As expected, the mean age in peptic ulcer diseases (PUD) patients was not significantly higher than in non ulcer dyspepsia (NUD) group (P=0.9). Demographic factors such as gender and education did not show any statistical differences in PUD and NUD.

Overall, 24 (72.7%) patients were infected with cagA-positive strains (Table 2). The mean age of these patients was 8.33 (\pm 1.87) years. The cagA gene was present in PUD and NUD patients, in 50% and 74.2% of strains, respectively. There were also no statistical differences between the cagA status and endoscopic or pathologic and clinical presentations irrespective of the peptic disease. This study showed that the presence of cagA gene was independent of the PUD risk,

Rafeey M, et al 553

Table 1: Demographic characteristics of patients, endoscopic and histopathology findings (n=106)

Variables	Levels	Frequency (%)
Gender	Male	52 (49.1)
	Female	54 (50.9)
Endoscopic finding	Ulcer	4 (3.8)
	Erythema	104 (98.1)
	Congestion	10 (9.4)
	Erosion	8 (7.5)
	Nodular gastritis	53 (50)
Pathologic finding	Acute gastritis	15 (14.2)
	Chronic gastritis	81 (76.4)
	Duodenitis	82 (77.4)
	Atrophy	3 (2.8)
	Esophagitis	95 (89.6)
H. pylori density	Negative	62 (58.5)
	Mild positive	39 (36.8)
	Moderate positive	4 (3.8)
	Severe positive	1 (0.9)
Chief complaint	Recurrent epigastric pain	88 (83)
	Vomiting	24 (22.6)
	Chronic diarrhea	5 (4.7)
	Failure to thrive	22 (20.8)
	GI bleeding	8 (7.5)

adjusted by age (P=0.8), sex (P=0.2) and other demographic data.

Thirty-three (100%) *H. pylori* isolates were carrying *vacA* gene. Genotypes of *vacA*, s1m2, s1m1, s2m2, and s2m1 were 45.5%, 30.3%, 21.2%

and 3%, respectively. There were also no statistical differences between the *vacA* genotypes and clinical outcomes both by univariate analyses and when adjusted by age, sex and other demographic data (Table 3). The *vacA*-m1 strains

Table 2: Relation between cagA and endoscopic and pathologic findings (n=24)

Variables	Level	cagA+ (%)	<i>P</i> -value	
Gender	Male	10 (41.7)	0.2	
	Female	14 (58.3)	0.2	
	Congestion	1 (4.2)	0.5	
	Erosion	2 (8.3)	0.4	
Endoscopic finding	Ulcer	1 (4.2)	0.4	
	Erythema	2 (100)	0.09	
	Nodular gastritis	15 (62.5)	0.7	
Pathologic finding	Chronic gastritis	16 (66.7)	0.5	
	Acute gastritis	7 (29.2)	0.4	
	Duodenitis	16 (66.7)	-	
	Esophagitis	22 (91.7)	0.8	
	Atrophy	0 (0)	0.9	
H. pylori density	Negative	9 (37.5)		
	Mild positive	14 (58.3)	0.4	
	Moderate positive	1 (4.2)	0.4	
	Severe positive	0 (0)		
Chief complaint	Epigastric pain	22 (91.7)	0.3	
	Vomiting	8 (33.3)	0.2	
	Chronic diarrhea	0 (0)	0.09	
	Failure to thrive	2 (8.3)	0.01	
	GI bleeding	3 (12.5)	0.22	

Variables	Level	S1 (%)	S2 (%)	M1 (%)	M2 (%)	P. value
Age(years)	Mean(SE)	9.36 (1.21)	8.09 (1.09)	9.63 (1.89)	8.72 (1.45)	0.5
Gender	Male	8 (36.4)	8 (72.7)	1 (12.5)	15 (60)	0.01
	Female	14 (63.6)	3 (27.3)	7 (87.5)	10 (40)	0.01
Endoscopic finding	Congestion	0 (0)	1 (9.1)	0 (0)	1 (4)	0.5
	Erosion	2 (9.1)	0 (0)	1 (12.5)	1 (4)	0.4
	Ulcer	1 (4.5)	1 (9.1)	0 (0)	2 (8)	0.4
	Erythema	21 (95.5)	11 (100)	8 (100)	24 (96)	0.5
	Nodular gastritis	13 (59.1)	7 (63.6)	7 (87.5)	13 (52)	0.07
Pathologic finding	Chronic gastritis	15 (68.6)	6 (54.5)	6 (75)	15 (60)	0.4
	Acute gastritis	6 (27.3)	5 (45.5)	2 (25)	9 (36)	0.6
	Duodenitis	13 (59.1)	9 (81.8)	6 (75)	16 (64)	0.6
	Esophagitis	19 (86.4)	11 (100)	7 (87.5)	23 (92)	0.7
	Atrophy	0 (0)	1 (9.1)	0 (0)	1 (4)	0.6
H. pylori density	Negative	9 (40.9)	3 (27.3)	4 (50)	8 (32)	
	Mild positive	12 (54.5)	7 (63.6)	4 (50)	15 (60)	0.8
	Moderate positive	0 (0)	1 (9.1)	0 (0)	1 (4)	
	Severe positive	1 (4.5)	0 (0)	0 (0)	1 (4)	
Chief complaint	Epigastric pain	18 (81.8)	11 (100)	8 (100)	21 (84)	0.2
	Vomiting	8 (36.4)	1 (9.1)	6 (75)	3 (12)	< 0.001
	Chronic diarrhea	1 (4.5)	0 (0)	0 (0)	1 (4)	0.6
	Failure to thrive	4 (18.2)	2 (18.2)	1 (12.5)	5 (20)	0.6
	GI bleeding	2 (9.1)	1 (9.1)	0 (0)	3 (12)	0.3
cagA		19 (80.4)	5 (45.5)	7 (87.5)	17 (68)	0.7

Table 3: Comparison of demographic data and vacA subgroups

were rather significantly more common (P=0.07) in patients with nodular gastritis. In this study s1m2 vacA genotype was more frequently detected in females than in males.

Discussion

There is ongoing interest in identifying *H. pylori* virulence factors that might predict the risk for clinical presentation. It has been proposed that *cagA* genes are such markers and can identify patients with peptic ulcer^[15].

The present study investigated the *cagA* and *vacA* genotypes of *H. pylori* isolated from pediatric population living in Azerbaijan, Iran. Because the strains were attained from symptomatic patients, the results reproduce the findings in these groups of patients rather than in the whole population. The current study confirms the distinctive difference in *H. pylori* genotypes in Iranian pediatric groups residing within the same region.

In some researches, the prevalence of H. pylori was reported to vary among different countries, regions, and patient groups, and it was reported as 82% in Brazil^[17], 78 to 80% in Turkey^[18,19], 82% in Japan^[20]. The majority of these studies indicated that in patients with duodenal ulcer, the cagA positivity rate is relatively higher than in patients with gastritis or gastric ulcer and ranges from 80 to 100%^[17-20]. In another study, the *cagA* positivity in Iranian isolates has been reported to vary from 44% to $91\%^{[11,21-23]}$. In the present study, 72.7% of the patients were infected with cagA positive strains; similar to other Iranian reports^[21,22,24,25]. However, this is different from studies in East and South Asian countries where more than 90% of the strains have the cagA gene irrespective of clinical outcome^[26-28]. Excitingly, many authors^{[29,} ^{30]} have found a significant correlation between the severity of histological alterations and the presence of the cagA gene in the H. pylori genome, whereas others[22,25,31,32] have been unable to support this relationship. The present study did not reveal associations between the cagA status and clinical presentation.

Rafeey M, et al 555

Our results revealed that cagA and vacA subtype m1 *H. pylori* strains were especially associated with nodular gastritis in Azerbaijan province. Hosseini et al found no correlation of caqA genotype and disease status, whereas vacA was demonstrated as a useful marker in predicting the disease outcome^[33]. van Doorn et al examined 94 gastric biopsy samples from patients in the Netherlands and reported that *cagA* positivity and vacA s1 genotype were associated with peptic ulcer disease^[15]. The present study did not reveal any association between the vacA and cagA status and PUD. This finding is in agreement with some reports from Iran^[13], but was different from other Iranian studies[25,34] and many studies from Western countries where vacA s1 and cagApositive strains are more often isolated from patients with PUD than with NUD[30]. In our study the number of patients is relatively small, so it is necessary that additional studies with large numbers of samples be studied to clarify the role of cagA, vacA in clinical presentation.

Conclusion

In conclusion, cagA and vacA-s1m1 genotypes are the predominant genotypes of H. pylori isolated from the northwestern Iranian pediatric population. However, we could not reveal clear association of cagA, and vacA genotypes with clinical presentation in pediatric age groups living in Azerbaijan province.

Acknowledgment

Conflict of Interest: None

References

 Milani, M, Ghotaslou R, Akhi MT, et al. The status of antimicrobial resistance of *Helicobacter pylori* in Eastern Azerbaijan, Iran: comparative study according to demographics. *J Infect Chemother* 2012; 18(6):848-52.

- Jabbari Moghaddam Y, Rafeey M, Radfar R. Comparative assessment of Helicobacter pylori colonization in children tonsillar tissues. Int J Pediatr Otorhinolaryngol 2009;73(9):1199-201.
- 3. Iranikhah A, Ghadir MR, Sarkeshikian S. Stool antigen tests for the detection of *Helicobacter Pylori* in children. *Iran J Pediatr* 2013;23(2):138-42.
- 4. Soltani J, Amirzadeh J, Nahedi S, et al. Prevalence of Helicobacter pylori infection in children, a population-based cross-sectional study in west of Iran. *Iran J Pediatr* 2013;23(1):13-8.
- 5. Rafeey M, Shabestari MS, Rafiey A, et al. The survey of Helicobacter pylori infection in infant. *Pak J Biol Sci* 2010;13(9):460-2.
- Salih BA. Helicobacter pylori infection in developing countries: The burden for how long? Saudi J Gastroenterol 2009;15(3):201-7.
- Malaty HM, Engstrand L, Pedersen NL, et al. Helicobacter pylori infection: genetic and environmental influences. A study of twins. *Ann Intern Med* 1994;120(12):982-6.
- 8. Graham DY. *Helicobacter pylori* infection in the pathogenesis of duodenal ulcer and gastric cancer: a model. *Gastroenterology* 1997;113(6):1983-91.
- Censini S, Lange C, Xiang Z, et al. Cag, a pathogenicity island of *Helicobacter pylori*, encodes type I-specific and disease-associated virulence factors. *Proc Natl Acad Sci USA* 1996;93(25):14648-53.
- 10. Yamaoka Y, Kita M, Kodama T, et al. Induction of various cytokines and development of severe mucosal inflammation by cagA gene positive Helicobacter pylori strains. Gut 1997;41(4):442-51.
- Atherton JC, Peek RM Jr, Tham KT, et al. Clinical and pathological importance of heterogeneity in *vacA*, the vacuolating cytotoxin gene of Helicobacter pylori. *Gastroenterology* 1997;112(1):92-9.
- 12. Jafari F, Shokrzadeh L, Dabiri H, et al. vacA genotypes of Helicobacter pylori in relation to cagA status and clinical outcomes in Iranian populations. *Jpn J Infect Dis* 2008;61(4):290-3.
- 13. Hussein NR, Mohammadi M, Talebkhan Y, et al. Differences in virulence markers between Helicobacter pylori strains from Iraq and those from Iran: potential importance of regional differences in H. pylori-associated disease. *J Clin Microbiol* 2008; 46(5):1774-9.
- 14. Dixon MF, Genta RM, Yardley JH, et al. Classification and grading of gastritis. The updated Sydney System. International Workshop on the Histopathology of Gastritis, Houston. *Am J Surg Pathol* 1996;20(10):1161-81.
- 15. van Doorn LJ, Figured C, Sanna R, et al. Clinical relevance of the *cagA*, *vacA*, and *iceA* status of Helicobacter pylori. *Gastroenterology* 1998;115(1): 58-66.
- 16. Sambrook J, Russell DW. Molecular Cloning: A Laboratory Manual. New York: CSHL press; 2001.

- 17. Ashour, AA, Magalhaes PP, Mendes EN, et al. Distribution of vacA genotypes in Helicobacter pylori strains isolated from Brazilian adult patients with gastritis, duodenal ulcer or gastric carcinoma. FEMS Immunol Med Microbiol 2002;33(3):173-8.
- 18. Bolek BK, Salih BA, Sander E. Genotyping of Helicobacter pylori strains from gastric biopsies by multiplex polymerase chain reaction. How advantageous is it? Diagn Microbiol Infect Dis 2007; 58(1):67-70.
- Saribasak H, Salih BA, Yamaoka Y, et al. Analysis of Helicobacter pylori genotypes and correlation with clinical outcome in Turkey. J Clin Microbiol 2004; 42(4):1648-51.
- 20. Takata T, Fujimoto S, Anzai K, et al. Analysis of the expression of CagA and VacA and the vacuolating activity in 167 isolates from patients with either peptic ulcers or non-ulcer dyspepsia. Am J Gastroenterol 1998;93(1):30-4.
- 21. Talebkhan Y, Mohammadi M, Mohagheghi MA. *cagA* gene and protein status among Iranian Helicobacter pylori strains. *Dig Dis Sci* 2008;53(4):925-32.
- 22. Molaei M, Foroughi F, Mashayekhi R, et al. CagA status and VacA subtypes of Helicobacter pylori in relation to histopathologic findings in Iranian population. *Indian J Pathol Microbiol* 2010;53(1): 24-7
- Ghasemian Safaei H, Tavakkoli H, Mojtahedi A, et al. Correlation of cagA positive Helicobacter pylori Infection with clinical outcomes in Alzahra hospital, Isfahan, Iran. J Res Med Sci 2008;13(4):196-201.
- Jafarzadeh A, Rezayati MT, Nemati M. Specific serum immunoglobulin G to H. pylori and CagA in healthy children and adults [south-east of Iran]. World J Gastroenterol 2007;13(22):3117-21.
- 25. Dabiri H, Bolfion M, Mirsalehian A, et al. Analysis of *Helicobacter pylori* genotypes in Afghani and Iranian isolates. *Pol J Microbiol* 2010;59(1):61-6.

- 26. Tan HJ, Rizal AM, Rosmadi MY, et al. Distribution of Helicobacter pylori cagA, cagE and vacA in different ethnic groups in Kuala Lumpur, Malaysia. J Gastroenterol Hepatol 2005;20(4):589-94.
- 27. Wong BC, Yin Y, Berg DE. Distribution of distinct vacA, cagA and iceA alleles in Helicobacter pylori in Hong Kong. *Helicobacter* 2001;6(4):317-24.
- Datta S, Chattopadhyay S, Balakrish Nair G. Virulence genes and neutral DNA markers of Helicobacter pylori isolates from different ethnic communities of West Bengal, India. J Clin Microbiol 2003;41(8):3737-43.
- 29. Korzon M, Sikorska-Wisniewska G, Jankowski Z. Clinical and pathological importance of cagApositive Helicobacter pylori strains in children with abdominal complaints. Helicobacter 1999;4(4):238-42.
- Queiroz DM, Mendes EN, Carvalho AS. Factors associated with *Helicobacter pylori* infection by a cagA-positive strain in children. *J Infect Dis* 2000; 181(2):626-30.
- 31. Yamaoka Y, Kodama T, Gutierrez O. Relationship between *Helicobacter pylori* iceA, *cag*A, and vacA status and clinical outcome: studies in four different countries. *J Clin Microbiol* 1999;37(7):2274-9.
- Gold BD, van Doorn LJ, Guarner J, et al. Genotypic, clinical, and demographic characteristics of children infected with Helicobacter pylori. *J Clin Microbiol* 2001;39(4):1348-52.
- Hosseini E, Poursina F, de Wiele TV, et al. Helicobacter pylori in Iran: A systematic review on the association of genotypes and gastroduodenal diseases. J Res Med Sci 2012;17(3):280-92.
- 34. Salari MH, Shirazi MH, Hadaiti MA, Daryani NA. Frequency of Helicobacter pylori vacA genotypes in Iranian patients with gastric and duodenal ulcer. *J Infect Public Health* 2009;2(4):204-8.