Comparison of a fluorometric assay kit with high-performance liquid chromatography for the assessment of serum retinol concentration.

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Abstract

Background: Although high-performance liquid chromatography (HPLC) is the commonly used method for the analysis of retinol in biological samples, simple and rapid test kits are available.

Objectives: This study compared a rapid test kit (ICHECK Fluoro®) to HPLC for the assessment of serum retinol concentrations.

Methods: For the analysis by HPLC, sample preparation included standard deproteinization and extraction phases. The analysis by ICHECK was performed by injecting serum into IEX reagent vials (n=89) and mixing manually for separation. After precipitation of the proteins, the vial was introduced into the chamber of the ICHECK Fluoro and analysed at 0 min (ICHECK0min) and 15 min later (ICHECK15min). Bland and Altman approach was applied to test the agreement between HPLC and ICHECK.

Results: Mean HPLC, ICHECK0min and ICHECK15min values were 421.2±106.0 µg/L, 423.1±118.3 µg/L and 413.2±107.6 µg/L, respectively. Retinol concentrations significantly decreased in the IEX solution over time (p<0.001). No significant proportional bias was observed between HPLC and ICHECK0min (r=-0.038, p=0.73) and ICHECK15min (r=-0.024, p=0.82). Fixed biases (HPLC minus ICHECK) for ICHECK0min and ICHECK15min were respectively -1.9±23.1 µg/l (p=0.45) and 8.0±22.7 µg/l (p=0.002).

Conclusion: ICHECK Fluoro may offer a reliable mean for assessing serum retinol for measurements performed with no significant time delay.

Keywords: HPLC, ICHECK Fluoro, serum retinol, test kit, vitamin A status.

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Introduction

Vitamin A deficiency (VAD) is a major health issue worldwide with significant impact on the disease burden¹-³. VAD is the leading cause for preventable sight-related diseases mainly xerophthalmia and blindness, and to the depletion of the immune function, increasing the risk of morbidity and mortality especially in children and pregnant women⁴,⁵. The prevalence of VAD has tremendously decreased worldwide, due to nutrition interventions including food fortification, mandatory supplementation, dietary diversification implementation, and nutritional education⁶,⁷. However, about 190 million children and 19.1 million pregnant women are still estimated as deficient in vitamin A, most of them are living in Africa and South-East Asia⁸.

The methods used to assess vitamin A status include clinical signs, serum retinol assessment, dose-response tests, and labelled isotopes; some of these methods even permit liver stores estimation⁹. At the population level, serum retinol is the most commonly recommended indicator to assess vitamin A status¹⁰,¹¹ and high-performance liquid chromatography (HPLC) is the most widely used technique for measuring retinol concentrations in serum samples¹². Thus, assessment of vitamin A status during and after nutritional interventions is routinely performed by measuring serum or plasma retinol levels.

ICHECK Fluoro® is a recent kit that can be used for the analysis of vitamin A concentrations in both fort-
The aim of our study was to evaluate the validity of the ICHECK for assessing serum retinol concentrations, comparatively to HPLC. Secondly, we hypothesized that working conditions in the field may result in some delay in the time of analysis. Therefore, we examined the evolution of retinol in the analysing solutions after a short period of time to assess the level of conservation of the retinol in the analysis solutions.

Methods
Serum samples were obtained from participants (n=89) of the PEN project, which is a one-year longitudinal study on vitamins A and D, iron and iodine deficiencies amongst school-age children (7 to 9 years) of a rural region in Morocco. Serum samples were stored in sealed 2.5 ml Eppendorf tubes at -80°C for less than 1 month before analysis. All the children and their parents had been clearly instructed about the protocol of the study and they provided a co-signed consent form. The PEN project was conducted under the ethical approval of the Ministry of Education of Morocco.

Chemicals
Ethanol, hexane, methanol, acetonitrile, retinol, retinyl acetate and butylated hydroxytoluene (BHT) of HPLC grade were purchased from Sigma-Aldrich (St. Louis, MO, USA). IEX Milia extraction kit was purchased from Bioanalyt (Potsdam, Germany).

HPLC
The HPLC consisted of a Waters system (Waters, Milford, MA, USA), a 2695 separation module, equipped with a 2996 PDA detector, a precolumn and a column C18 Sunfire, 5µm, 4.6x250 mm. Serum samples were thawed at room temperature for 15 minutes. A volume of 500 µl of serum was added to the IEX solution via using 1ml syringe. The IEX vial was held between the thumb and the index fingers and vigorously mixed for 10 seconds. After precipitation of the proteins, the IEX vial was introduced into the IEX analysing chamber and assayed at time 0 (ICHECK0min) and after 15 min (ICHECK15min). Calibration of the ICHECK device was performed using a sealed calibration solution prepared on the day of analysis. Retinol was dissolved in ethanol and used as external standard. All the analysis was performed under yellow-orange light to prevent the degradation of retinoids.

ICHECK
The ICHECK Fluoro device was manufactured by Bioanalyt (Potsdam, Germany). A set of 96 analyzing IEX vials was used: 89 were analysed, 5 were used for control and 2 were discarded. Serum samples were allowed to thaw at room temperature for 15 minutes. Then, a volume of 500 µl of serum was added to the IEX solution via using 1ml syringe. The IEX vial was held between the thumb and the index fingers and vigorously mixed for 10 seconds. After precipitation of the proteins, the IEX vial was introduced into the ICHECK chamber and assayed at time 0 (ICHECK0min) and after 15 min (ICHECK15min). Calibration of the ICHECK device was performed using a sealed calibration solution provided by the manufacturer.

Statistical analysis
Data was analysed using STATA 12 (StataCorp, College Station, TX, USA). Descriptive statistics were presented as mean±standard deviation (SD). The relation between HPLC and ICHECK measures was determined by the correlation of Pearson. Agreement between HPLC and ICHECK measures was examined using Bland and Altman analysis. Fixed biases (HPLC minus ICHECK value) and ICHECK0min was not significant (1.9±23.2 µg/L, p=0.45) with a confidence interval (CI) at 95 % ranging from -7.0 µg/L to 3.1 µg/L whereas it was significant between HPLC and ICHECK15min (8.0±22.7 µg/L, p=0.002, CI : 3.1 to 13.4 µg/L).

Results
Serum retinol levels measured by HPLC and ICHECK are presented in Table 1. Agreement analysis between HPLC and ICHECK measures is presented in Table 2. The correlation coefficients between HPLC, ICHECK0min and ICHECK15min were significant. The fixed bias between HPLC and ICHECK0min was not significant (1.9±23.2 µg/L, p=0.45) with a confidence interval (CI) at 95 % ranging from -7.0 µg/L to 3.1 µg/L whereas it was significant between HPLC and ICHECK15min (8.0±22.7 µg/L, p=0.002, CI : 3.1 to 13.4 µg/L).

Table 1. Serum retinol by HPLC and ICHECK

<table>
<thead>
<tr>
<th></th>
<th>Mean (µg/L)</th>
<th>SD</th>
<th>Range</th>
</tr>
</thead>
<tbody>
<tr>
<td>HPLC</td>
<td>421.2</td>
<td>106.0</td>
<td>110 - 695</td>
</tr>
<tr>
<td>ICHECK</td>
<td>423.1</td>
<td>108.3</td>
<td>108 - 746</td>
</tr>
<tr>
<td>ICHECK15min</td>
<td>413.2</td>
<td>107.6</td>
<td>105 - 730</td>
</tr>
</tbody>
</table>

Table 2. Comparison between serum retinol determined by HPLC and ICHECK

<table>
<thead>
<tr>
<th></th>
<th>Correlation</th>
<th>Fixed bias± SD</th>
<th>p-value</th>
<th>Limits of agreement</th>
<th>Proportional bias</th>
</tr>
</thead>
<tbody>
<tr>
<td>HPLC - ICHECK0min</td>
<td>0.977</td>
<td>-1.9±23.2</td>
<td>0.45</td>
<td>-48.2; 44.3</td>
<td>-0.038 (p=0.73)</td>
</tr>
<tr>
<td>HPLC - ICHECK15min</td>
<td>0.978</td>
<td>8.0±22.7</td>
<td>0.002</td>
<td>-37.5; 53.4</td>
<td>-0.025 (p=0.83)</td>
</tr>
<tr>
<td>ICHECK0min - ICHECK15min</td>
<td>0.995</td>
<td>-9.9±10.5</td>
<td>&lt;0.00</td>
<td>-30.9; 11.1</td>
<td>-0.092 (p=0.40)</td>
</tr>
</tbody>
</table>

SD, standard deviation. Vitamin A values are in µg/L.
*Pearson correlation coefficient
bHPLC minus ICHECK value, or ICHECK15min minus ICHECK0min/a negative value reflects an overestimation
cStudent t-test value for the equality of the difference to 0
#Mean±2SD
**Pitman’s test

There was a decrease of the concentration of retinol in the IEX analysing solution over time. This was showed by a significant fixed bias between ICHECK0min and ICHECK15min (<9.9±10.5 µg/L, p<0.001). None of the values of the ICHECK showed a significant proportional bias with HPLC. Bland and Altman plots between HPLC and ICHECK0min and ICHECK15min are presented respectively in Figure 1a and Figure 1b.
The determination of vitamin A status through the assessment of liver reserves, which is considered the gold standard method, is difficult to apply when samples are large\(^1\). Thus, serum retinol measurement represents a valuable alternative. However, intra- and inter-individual variations of serum retinol suggest that the interpretation of serum retinol levels is only valid in populations, and is useless at individual levels\(^1\). Moreover, vitamin A is sensitive to light, heat and oxidation, and losses can occur during sample collection, centrifugation, transport and storage.

ICHECK Fluoro is a fluorometry-based portable kit, easy-to-use and not requiring highly trained technicians. The ICHECK briefcase includes a digital mini-scale for solid samples, a multiple charger adapted to remote areas, a calibration solution, additionally to IEX Mila solutions and the ICHECK device. We were concerned about the stability of retinol in the IEX solution over a period of time. We assumed that 15 minutes might represent the average interval of time during which an overloaded technician would keep the samples before analysis.

Our result showed that ICHECK measurement provided good correlations with the HPLC for serum retinol concentrations. Nevertheless, our study showed that the concentration of retinol decreased in the IEX solution over time. As expected, assay performed directly after separation of the IEX-serum complex provided better agreement with HPLC. The degradation of retinol in organic solvents offers a plausible explanation for the decrease of retinol concentrations over time. Several authors reported that the decrease of retinol concentrations started immediately after addition of organic solvents to serum even at ice temperature\(^1\). Previous studies reported that addition of an antioxidant, especially ascorbic acid\(^2\), butylated hydroxyanisole (BHA) and BHT\(^3\), to solvents during extraction slowed the degradation of retinol.

The protocol of our study did not include the addition of an antioxidant to the serum or to the IEX kit to test the hypothesis of conservation. Thus, this can be considered as a limitation of the study. Another limitation of this study was the low prevalence of vitamin A deficiency among participants (only two participants). However, the non-significance of the proportional bias showed that the device may be suitable to provide appropriate measurement of retinol concentrations at both lower and higher concentrations.

The concentration of serum retinol in IEX reagent solution decreased over time, therefore the complex IEX-serum sample should be analysed immediately after the separation of the supernatant. Otherwise, further studies might test the effect of the addition of appropriate antioxidants in the IEX solution. Overall, based on the simplicity of the analysis procedure, ICHECK device should be recommended for field and epidemiologic studies, especially in developing countries.

**Conflict of interest**
None of the authors declared a conflict of interest.

**Acknowledgements**
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**References**